

## Microcrystalline Cellulose

### Add the following:

▲Portions of this monograph that are national *USP* text, and are not part of the harmonized text, are marked with symbols (▲, ◆) to specify this fact. ▲ NF 1-Dec-2019

Cellulose [9004-34-6].

### DEFINITION

Microcrystalline Cellulose is purified, partially depolymerized cellulose prepared by treating alpha cellulose, obtained as a pulp from fibrous plant material, with mineral acids.

### IDENTIFICATION

#### Add the following:

▲[NOTE—Compliance is determined by meeting the requirements of *Identification A, B, and C.*]▲ NF 1-Dec-2019

#### Add the following:

- ▲ **A. INFRARED ABSORPTION** (197K) or (197A):  
[NOTE—Disregard any peak between 800 and 825  $\text{cm}^{-1}$  as well as those between 950 and 1000  $\text{cm}^{-1}$ .]▲ NF 1-Dec-2019

#### Change to read:

- ▲ **B.**▲ NF 1-Dec-2019  
**Iodinated zinc chloride solution:** Dissolve 20 g of zinc chloride and 6.5 g of potassium iodide in 10.5 mL of water. Add 0.5 g of iodine, and shake for 15 min.  
**Sample:** 10 mg  
**Analysis:** Place the *Sample* on a watch glass, and disperse in 2 mL of *Iodinated zinc chloride solution*.  
**Acceptance criteria:** The substance takes on a violet-blue color.

#### Change to read:

- ▲ **C.**▲ NF 1-Dec-2019  
**Sample:** 1.3 g of Microcrystalline Cellulose, accurately weighed to 0.1 mg  
**Analysis:** Transfer the *Sample* to a 125-mL conical flask. Add 25.0 mL of water and 25.0 mL of 1.0 M cupriethylenediamine hydroxide solution. Immediately purge the solution with nitrogen, insert the stopper, and shake on a wrist-action shaker, or other suitable mechanical shaker, until completely dissolved. Transfer an appropriate volume of the sample solution to a calibrated number 150 Cannon-Fenske, or equivalent, viscometer. Allow the solution to equilibrate at  $25 \pm 0.1^\circ$  for NLT 5 min. Time the flow between the 2 marks on the viscometer, and record the flow time,  $t_1$ , in seconds. Calculate the kinematic viscosity,  $(KV)_1$ , of Microcrystalline Cellulose taken:

$$\text{Result} = t_1 \times k_1$$

- $t_1$  = flow time (s)
- $k_1$  = viscometer constant (see *Viscosity—Capillary Methods* (911))

Obtain the flow time,  $t_2$ , for 0.5 M cupriethylenediamine hydroxide solutions using a number 100 Cannon-Fenske, or equivalent, viscometer.  
Calculate the kinematic viscosity,  $(KV)_2$ , of the solvent:

$$\text{Result} = t_2 \times k_2$$

- $t_2$  = flow time for 0.5 M cupriethylenediamine hydroxide solutions (s)
- $k_2$  = viscometer constant

Determine the relative viscosity,  $\eta_{rel}$ , of the Microcrystalline Cellulose specimen taken:

$$\text{Result} = (KV)_1 / (KV)_2$$

- $(KV)_1$  = kinematic viscosity of Microcrystalline Cellulose taken
- $(KV)_2$  = kinematic viscosity of the solvent

Determine the intrinsic viscosity,  $[\eta]_c$ , by interpolation, using the *Intrinsic Viscosity Table* in the *Reference Tables* section.

Calculate the degree of polymerization,  $P$ :

$$\text{Result} = [(95) \times [\eta]_c / \{W_5 \times [(100 - \% \text{LOD}) / 100]\}]$$

- $[\eta]_c$  = intrinsic viscosity
- $W_5$  = weight of Microcrystalline Cellulose taken (g)
- $\% \text{LOD}$  = value obtained from the test for *Loss on Drying*

**Acceptance criteria:** The degree of polymerization is NMT 350.

### IMPURITIES

#### INORGANIC IMPURITIES

- **Residue on Ignition** (281): NMT 0.1%

#### SPECIFIC TESTS

- **MICROBIAL ENUMERATION TESTS** (61) and **TESTS FOR SPECIFIED MICROORGANISMS** (62): The total aerobic microbial count does not exceed  $10^3$  cfu/g, and the total combined molds and yeasts count does not exceed  $10^2$  cfu/g. It meets the requirements of the tests for absence of *Staphylococcus aureus* and *Pseudomonas aeruginosa* and for the absence of *Escherichia coli* and *Salmonella species*.
- **CONDUCTIVITY**  
**Sample:** 5 g  
**Analysis:** Shake the *Sample* with 40 mL of water for 20 min, and centrifuge. Retain the supernatant for use in the *pH* test. Using an appropriate conductivity meter that has been standardized with a potassium chloride conductivity calibration standard having a conductivity of 100  $\mu\text{S}/\text{cm}$ , measure the conductivity of the supernatant after a stable reading is obtained, and measure the conductivity of the water used to prepare the test specimen.  
**Acceptance criteria:** The conductivity of the supernatant does not exceed the conductivity of the water by more than 75  $\mu\text{S}/\text{cm}$ .
- **pH** (791): 5.0–7.5 in the supernatant obtained in the *Conductivity* test

#### Change to read:

- **LOSS ON DRYING** (731)  
**Analysis:** Dry a sample at  $105^\circ$  for 3 h.  
**Acceptance criteria:** NMT 7.0%▲◆ NF 1-Dec-2019 or some other lower percentage, or is within a percentage range, as specified in the labeling▲◆ NF 1-Dec-2019
- **BULK DENSITY**  
**Analysis:** Use a volumeter that has been fitted with a 10-mesh screen. The volumeter is freestanding of the brass or stainless steel cup, which is calibrated to a capacity of

25.0 ± 0.05 mL and has an inside diameter of 30.0 ± 2.0 mm. Weigh the empty cup, position it under the chute, and slowly pour the powder from a height of 5.1 cm (2 in) above the funnel through the volumeter, at a rate suitable to prevent clogging, until the cup overflows.

[NOTE—If excessive clogging of the screen occurs, remove the screen.] Level the excess powder, and weigh the filled cup. Calculate the bulk density by dividing the weight of the powder in the cup by the volume of the cup.

**Acceptance criteria:** The bulk density is within the labeled specification.

#### Change to read:

- ▲▲ NF 1-Dec-2019 **PARTICLE SIZE DISTRIBUTION**

[NOTE—In cases where there are no functionality-related concerns regarding the particle size distribution of the article, this test may be omitted.]

Where the labeling states the particle size distribution, determine the particle size distribution as directed in *Particle Size Distribution Estimation by Analytical Sieving* (786), or by a suitable validated procedure. ▲▲ NF 1-Dec-2019

- **WATER-SOLUBLE SUBSTANCES**

**Sample:** 5.0 g

**Analysis:** Shake the *Sample* with 80 mL of water for 10 min, and pass with the aid of a vacuum through filter paper (Whatman No. 42 or equivalent) into a vacuum flask. Transfer the filtrate to a tared beaker, evaporate to dryness without charring, dry at 105° for 1 h, cool in a desiccator, and weigh.

**Acceptance criteria:** The difference between the weight of the residue and the weight obtained from a blank determination does not exceed 12.5 mg (0.25%).

- **ETHER-SOLUBLE SUBSTANCES**

**Sample:** 10.0 g

**Analysis:** Place the *Sample* in a chromatographic column having an internal diameter of about 20 mm, and pass 50

mL of peroxide-free ether through the column. Evaporate the eluate to dryness in a previously dried and tared evaporating dish with the aid of a current of air in a fume hood. After all the ether has evaporated, dry the residue at 105° for 30 min, cool in a desiccator, and weigh.

**Acceptance criteria:** The difference between the weight of the residue and the weight obtained from a blank determination does not exceed 5.0 mg (0.05%).

#### ADDITIONAL REQUIREMENTS

##### Change to read:

- ▲▲ NF 1-Dec-2019 **PACKAGING AND STORAGE:** Preserve in tight containers. ▲▲ NF 1-Dec-2019

##### Change to read:

- ▲▲ NF 1-Dec-2019 **LABELING:** The labeling indicates the nominal loss on drying, bulk density, and degree of polymerization values. Degree of polymerization compliance is determined using *Identification* ▲C. ▲ NF 1-Dec-2019 Where the particle size distribution is stated in the labeling, proceed as directed in the test for *Particle Size Distribution*. The labeling indicates with which technique the particle size distribution was determined if a technique other than analytical sieving was used; and the labeling indicates the  $d_{10}$ ,  $d_{50}$ , and  $d_{90}$  values and the range for each. ▲▲ NF 1-Dec-2019

##### Add the following:

- ▲• **USP REFERENCE STANDARDS** (11)

USP Microcrystalline Cellulose RS

▲ NF 1-Dec-2019