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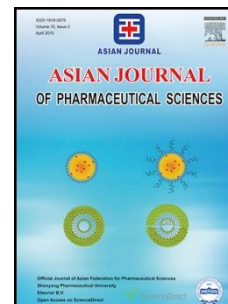
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Thin films as an emerging platform for drug delivery

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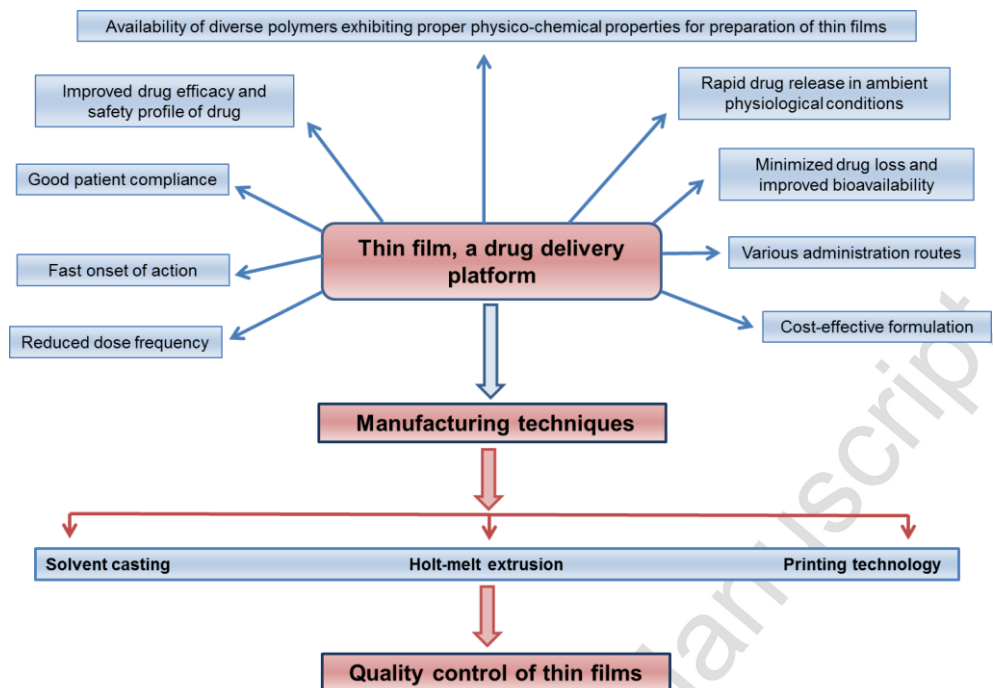
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27 **Graphical abstract**

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31 This review provides an overview of critical factors, characterization methods, and quality
 32 specifications for development of thin film formulations for drug delivery along with the
 33 recent trends and future perspectives.

34

35 **Abstract**

36

37 Pharmaceutical scientists throughout the world are trying to explore thin films as a novel
 38 drug delivery tool. Thin films have been identified as an alternative approach to conventional
 39 dosage forms. The thin films are considered to be convenient to swallow, self-administrable,
 40 and fast dissolving dosage form; all of which makes it as a versatile platform for drug
 41 delivery. This delivery system has been used for both systemic and local action *via* several
 42 routes such as oral, buccal, sublingual, ocular, and transdermal routes. The design of efficient
 43 thin films requires a comprehensive knowledge of the pharmacological and pharmaceutical
 44 properties of drugs and polymers along with an appropriate selection of manufacturing

45 processes. Therefore, the aim of this review is to provide an overview of the critical factors
46 affecting the formulation of thin films including the physico-chemical properties of polymers
47 and drugs, anatomical and physiological constraints, as well as the characterization methods
48 and quality specifications to circumvent the difficulties associated with formulation design. It
49 also highlights the recent trends and perspectives to develop thin film products by various
50 companies.

51

52 **Keywords:** Thin film, Film-forming polymer, Mechanical properties, Manufacturing,
53 Characterization

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56 1. Introduction

57 Generally, thin films can be referred as a thin and flexible layer of polymer with or without
58 a plasticizer [1]. Since they are thin and flexible by their nature, it can be perceived to be less
59 obtrusive and more acceptable by the patient [2]. The thin film is polymeric matrices that
60 meet many requirements for being used efficiently as a drug release platform [3].
61 Fundamentally, thin films are excellent candidates for targeting sensitive site that may not be
62 possible with tablets or liquid formulations [4]. Thin films have shown the capabilities to
63 improve the onset of drug action, reduce the dose frequency and enhance the drug efficacy
64 [3]. Similarly, thin films may be useful for eliminating side effects of a drug and reducing
65 extensive metabolism caused by proteolytic enzymes [5, 6]. Ideal thin films need to exhibit
66 desirable features such as sufficient drug loading capacity, fast dissolution rate or long
67 residence time at the site of administration, and acceptable formulation stability. They should
68 also be non-toxic, biocompatible and biodegradable [7, 8].

69 Compared with the existing traditional dosage forms, it stands out to be superior in terms
70 of enhanced bioavailability, high patient compliance, and patent extension of active
71 pharmaceutical ingredients (API) [9]. Furthermore, thin film formulations offer several
72 advantages including: (a) convenient administration through non-invasive routes, (b) ease of
73 handling during manufacture and transportation, and (c) cost-effectiveness in the
74 development of formulations [8, 10, 11]. The availability of a wide array of suitable polymers
75 and the paradigm shift in manufacturing technology have made possible to develop a wide
76 range of thin films [12]. Therefore, a thin film is gaining popularity and acceptance in the
77 pharmaceutical arena as a novel drug delivery dosage form.

78 Substantial efforts have been made to formulate polymeric thin films that are administered
79 generally *via* buccal, sublingual, ocular and skin routes [13, 14]. Among different routes, the
80 use of thin films for delivering medicine into sublingual or buccal mucosa has drawn
81 immense interest in recent years [15]. Meanwhile, ophthalmic films are currently developed
82 for overcoming the ocular barriers and preventing loss of drugs through the lacrimal drainage
83 system [16]. Controlling compositions of polymers of different grades has facilitated the
84 modification of key characteristics of thin films such as drug release rate, mucoadhesive
85 properties, mechanical strength and other related properties. Additionally, various inactive
86 components can be included such as fillers, plasticizer, saliva stimulating agent, colorants,
87 and sweeteners for improving aesthetic characteristics. Many pharmaceutical companies are

88 fascinated by the appealing features of thin films and as a result, they have already patented
89 various technologies for producing thin films [17].

90 Currently, a significant amount of original works and patents can be found in literature,
91 but, still there is a need for extensive studies to optimize the performance of thin films
92 accurately. The lack of appropriate guidance for the manufacture, characterization and quality
93 control of the thin films has sought the need of adequate studies in this area from the
94 pharmaceutical viewpoint. Therefore, this paper will contribute to give insights on
95 understanding the critical quality attributes and characterization methods with the aim to
96 enhance the performance of thin films.

97

98 **2. Types of thin films**

99

100 Thin film is not a recent formulation, and it was first introduced in late 1970 to overcome
101 swallowing difficulties exhibited by tablets and capsules [15]. Various names of thin films
102 are appeared such as oral film (oral thin film), oral soluble film, wafer, oral strip,
103 orodispersible film (ODF), buccal film, mucoadhesive film, ophthalmic film, and
104 transmucosal film. While several films are designed to be dissolved quickly in the oral cavity
105 for the absorption of a drug in the gastrointestinal cavity (oral and oral soluble or,
106 orodispersible films), some are prepared to deliver a drug at the site of administration (e.g.,
107 buccal, sublingual and ophthalmic thin films). Drugs with high mucosal permeability have
108 been known to be suitable for buccal and sublingual delivery with films [18]. Likewise,
109 ophthalmic thin films are generally applied to treat diseases of the anterior segment such as
110 conjunctivitis, glaucoma and chronic dry eye syndromes [5, 19].

111 A film that readily dissolves in the oral cavity is generally termed as orodispersible film
112 according to European Medicines Agency (EMA) or simply soluble film according to FDA
113 [3]. Usually, fast dissolving oral films are ultra-thin film (50-150 μm) having size of postage
114 stamp, which dissolves within a min in the oral cavity after being in contact with the saliva
115 resulting in quick absorption and instant bioavailability of the drugs [20, 21]. Drugs loaded in
116 buccal adhesive films are absorbed directly *via* buccal mucosa, which delivers the drug to the
117 systemic circulation after their absorption [22]. Likewise, wafer is frequently mentioned as
118 paper-thin polymeric films employed as carriers for pharmaceutical agents. This innovative
119 dosage form is taken orally but does not require water to swallow for the absorption of a drug
120 [23]. Orodispersible films should not be misunderstood with buccal films designed for

121 staying longer on the cheek mucosa [24]. Therefore, different types of films should be
122 distinguished accurately to prevent possible misinterpretations.

123

124 **3. Advantages of thin films as an emerging dosage form**

125

126 *3.1. Advantages over conventional dosage forms*

127

128 A thin film dissolves rapidly than other conventional dosage forms [25]. Thin films are less
129 friable and easy to carry dosage form compared to commercialized orally fast disintegrating
130 tablets, which need special packing. Likewise, a single dose of strip can be carried
131 individually without requiring the secondary container [26, 27]. It is very important to
132 address the poor stability of liquid dosage forms, especially the aqueous formulations. Unlike
133 the thin films, there is a need of great care during accurate measurement of the amount and
134 shaking the bottle every time before administration may contribute to less acceptance by the
135 patients [3]. Conventional ophthalmic drug delivery systems such as eye drops or solutions
136 are commonly used but they are limited in their ability to provide high ocular drug
137 bioavailability and sustained duration of action [28]. Ophthalmic thin films can be used to
138 improve the drug delivery to the eye. In contrast to transdermal patch, the transdermal film is
139 less associated with skin irritation due to less occlusive properties that improve the water
140 vapour permeation through the skin and do not leave sticky sensation on the site of
141 application [29, 30].

142

143 *3.2. Clinical advantages*

144

145 Patients show preference towards thin film due to its appellative form and ease of
146 administration [17]. Furthermore, oral dissolving film is extensively useful for pediatric,
147 geriatric, and psychiatric patients since it is easy to administer and avoid the risk of choking
148 or suffocation, thus ensuring patient safety [22]. Ophthalmic films have known to enhance
149 the retention time of a drug and thereby, the absorption of the drug was greatly improved
150 from the anterior segment of the eye [31]. Moreover, the polymeric thin films can also be
151 beneficial for bedridden and non-cooperative patients as they can be administered easily and
152 hardly spit out. A thin film is useful in cases where a rapid onset of action is required such as
153 in motion sickness, sudden episodes of allergic attack or coughing, bronchitis or asthma [22].

154

155

156 **4. Major limitations of thin films**

157

158 Use of thin films is sometimes limited largely due to low drug loading capacity for a less
159 potent drug given at high dose [10]. Thin films are usually hygroscopic in nature. Thus,
160 special precaution should be taken for their longer preservation [4]. Combining more than
161 one drug concomitantly is a very challenging task in oral film formulation because both the
162 dissolution rate as well as the disintegration time are hindered by the co-administration of a
163 drug in oral films [32]. The difficulty to obtain a high degree of accuracy with respect to the
164 amount of drug in individual unit dose of the film can lead to therapeutic failure, non-
165 reproducible effects and sometimes toxic effects to the patient [33]. Preparing oral film
166 formulation is concerned with the issues of requiring excessive time for drying. It takes
167 around one day for the complete drying at room temperature, which notably decrease the rate
168 of production of films. Since it is not recommended to use hot air oven for thermolabile
169 drugs, an alternative process of drying should be explored [22].

170

171

172 **5. Polymers for the preparation of thin films**

173

174 Polymers are the backbone of film formulations and various polymers are available for the
175 preparation of thin films [34]. The polymers can be used alone or in combination with other
176 polymers to achieve the desired film properties. The polymers employed should be non-toxic,
177 non-irritant, and absence of leachable impurities is required. Water-soluble polymers are used
178 as film formers to produce a thin film with rapid disintegration, good mechanical strength,
179 and good mouthfeel effects. Both natural and synthetic polymers are used for film preparation
180 [20, 35]. The list of polymers commonly used in the manufacture of polymeric films, with
181 additional descriptions and properties, is depicted in **Table 1**.

182 Availability of diverse polymers allows imparting specific properties in the thin films. For
183 instance, gelatins are available in different molecular weights, and thus, the appealing and
184 glossy films could be obtained with the gelatin having a high molecular weight. Pullulan is
185 frequently used for producing a thin film with great solubility, high mechanical strength and
186 they are stable over a wide range of temperatures. The blending of chitosan and high methoxy
187 pectin (HMP) or low methoxy pectin (LMP) resulted in a thin film exhibiting an excellent

188 mechanical strength. The film forming polymers such as hydroxypropyl cellulose (HPC),
189 methyl cellulose, and CMC produce a thin film with less water vapour barrier due to
190 hydrophilic nature which aids in water retention [15].

191 In one study, a fast-dissolving film of triclosan was prepared using different grades of
192 HPMC named as Methocel E3, Methocel E5, and Methocel E15 Premium LV as a primary
193 film former. The result demonstrated that Methocel E5 Premium LV at the concentration of
194 2.2% w/v produced films with excellent film properties [37]. The *in vitro* residence time of
195 the film made from Carbopol® 934P and HPMC E15 was almost double than the films
196 containing only HPMC E15. Additionally, it was observed that the combined polymers were
197 more resistance to breakage [11]. Cilurzo et al. (2008) reported the use of maltodextrins
198 (MDX) with low dextrose content as a film forming polymer for the preparation of oral fast-
199 dissolving films of an insoluble drug, piroxicam. Despite the decrease in film ductility due to
200 the loading of the drug as a powder, the produced film exhibited satisfactory flexibility and
201 resistance to elongation along with rapid dissolution [38]. Similarly, oral dissolving films of
202 granisetron HCl manufactured using HPMC and pullulan illustrated the effect of increasing
203 polymer concentration on mechanical properties and physical properties of films. Pullulan
204 with 40-45% concentration was not able to produce films with good strength whereas the
205 HPMC used in 40% concentration yielded the film which was difficult to peel. Likewise, the
206 film stickiness increased when the concentration of HPMC was beyond 50% [39].

207 Mucoadhesive films are thin and flexible retentive dosage forms, and release drug directly
208 into a biological substrate. They facilitate in extending residence time at the application site
209 leading to prolonged therapeutic effects [40]. Majority of the thin film having mucoadhesive
210 properties are hydrophilic in nature that undergoes swelling and form a chain interaction with
211 the mucin [11]. Among the several studied polymers, the most compelling mucoadhesion
212 properties are exhibited by chitosan, hyaluronan, cellulose derivatives, polyacrylates,
213 alginate, gelatin and pectin [41]. Compared with non-ionic polymers, the cationic and anionic
214 polymers facilitate strong interaction with mucus [42]. Anionic polymers are well-
215 characterized due to the existence of carboxyl and sulfate functional groups, which create the
216 negative charge at pH values surpassing the pKa of the polymer. As an example, sodium
217 carboxymethyl cellulose (NaCMC), and polyacrylic acid (PAA) exhibit excellent
218 mucoadhesive properties because of bond formation with the mucin [43]. Thiomers i.e.
219 polymer containing thiol group stand out to enhance mucoadhesion because they are able to
220 interact with the mucin through the formation of disulphide linkages. The process of
221 'thiolation' is possible with many polymers, using amide-coupling chemistry, where the

222 aqueous solvent systems are used [44]. Eudragit displayed promising mucoadhesive
223 properties when used alone or in combination with other hydrophilic polymers. Films,
224 prepared from the propranolol HCl, Eudragit RS100, and triethyl citrate (plasticizer),
225 demonstrated mucoadhesive force three times greater than the film prepared with chitosan as
226 the mucoadhesive polymer [11]. Juliano et al. (2008) prepared a buccoadhesive films
227 constituting alginate and/or HPMC and/or chitosan either as a single polymer or in a
228 combination of two. Basically, they aimed the films to release the chlorhexidine diacetate in a
229 controlled manner. HPMC was not able to prolong the chlorhexidine release as more than
230 80% of the drug was released within only 30 min. However, chlorohexidine incorporated in
231 alginate and alginate/chitosan-based films showed that only 30-35% of the drug was released
232 in 30 min; hence, this polymeric system is beneficial for prolonged drug release [45].

233 In common terms, polymers are understood as excipients, but it has become an essential
234 component while designing and formulating thin films. Therefore, understanding the
235 properties of polymers such as chemistry, rheology, physico-chemical properties of polymer
236 seems to be imminent for maximizing their uses to develop a thin film. The selection of
237 appropriate polymer during the development of polymeric thin films may be critical; thereby,
238 several points should be considered according to the requirements. Therefore, it is imperative
239 to consider the appropriate polymer for producing a thin film with a better performance that
240 assures high therapeutic success.

241

242 **6. Technologies for manufacturing thin films**

243

244 The most commonly used techniques for the preparation of thin films are solvent casting
245 [46, 47] and hot melt extrusion [38, 48]. However, an innovative technique like inkjet
246 printing [49] has evolved in the past few years. Various methods that have been employed for
247 polymeric thin film manufacturing are described below in detail:

248

249 *6.1. Solvent casting*

250

251 Among several techniques of film manufacturing, solvent casting is feasible, preferable
252 and undoubtedly widely used method mainly due to the straightforward manufacturing
253 process and low cost of processing. The manufacturing procedure of thin films with the
254 solvent casting method along with the quality control parameters in each step is illustrated in

255 **Fig. 1.** The rheological properties of the polymeric mixture should be taken into account
256 since they affect the drying rate, the film thickness, the morphology as well the content
257 uniformity of the films [26]. The mixing process could introduce the air bubbles into the
258 liquid inadvertently; therefore, de-aeration is a pre-requisite to obtain a homogeneous product
259 [17]. After casting the solution into a suitable substrate, they are left for drying to allow the
260 solvent to evaporate that just leaves a polymeric film with a drug on it [2].

261 After the complete drying of the film, it is cut into suitable shape and size depending upon
262 the required dosage of the formed strip. In the majority of the cases, the strips are rolled and
263 stored for a certain time before cutting, which is known as 'rollstock' in an industry.
264 However, a film should not be exposed for too long time since it is prone for being damaged.
265 If possible, it should be cut and packed immediately after the preparation to keep its stability
266 [17]. Several advantages such as better physical properties, easy and low cost processing, and
267 excellent uniformity of thickness are observed with the film obtained by solvent-casting [50].
268 However, this process suffers from some limitation. For instance, a polymeric thin film
269 prepared by solvent casting method was brittle upon storage, as marked by decrease in the
270 percent elongation due to evaporation or loss of the residual solvent in the film over time
271 [51]. Another issue under scrutiny associated with this method is the requirement of using
272 organic solvents. The presence of organic solvent system is a serious problem because it
273 causes a hazard to health and environment. As a result, strict regulations have been adopted
274 by many countries regarding the use of an organic solvent [11].

275 Translating the production of films from a bench scale to production scale is one of the
276 biggest challenges because many factors such as heating, mixing speed, and temperature
277 could bring variability in quality and consistent formation of films in commercial scale may
278 not be possible. Therefore, sufficient endeavor should be invested to optimize the various
279 parameters such as the speed of casting, drying time, and final thickness of the dried strip,
280 which may affect the production of films from commercial scale output [17]. **Fig. 2** depicts
281 the machine that is used for a large-scale production of film based on solvent casting
282 technique.

283

284 6.2. Hot-melt extrusion (HME)

285

286 HME is a versatile method adopted for the manufacture of granules, tablets, pellets [52],
287 and also thin films [38]. It is a substitute method to solvent casting for the preparation of the
288 film, especially useful when no organic solvent system is required [10]. However, only few

289 literature has reported the use of hot-melt extrusion for the preparation of polymeric thin
290 films [11]. HME is a process of shaping a mixture of polymers, drug substance, and other
291 excipients into a film by melting all the components [3]. Eventually, the films are cut into a
292 particular shape and dimensions [6]. In this method, a mixture of pharmaceutical ingredients
293 is molten and then charged through an orifice (the die) to obtain homogeneous matrices [11].
294 Since APIs are subjected to operation at high-temperature with complete absence of solvents,
295 this method is not suitable for thermos-labile APIs [17]. The practical steps of HME are
296 outlined as follows [53]:

- 297 (i) Feeding of the components to the extruder through a hopper,
298 (ii) Mixing, grinding, and kneading,
299 (iii) Flowing the molten and blended mass to the die, and
300 (iv) Extruding the mass through the die and further downstream processing

301 The equipment for the process of HME is illustrated in **Fig. 3**, which consists of the
302 hopper, extruder, film die, and roller. The extruder contains one or two rotating screws (co-
303 rotating or counter rotating) inside a static cylindrical barrel. The barrel is often manufactured
304 in sections to shorten the residence time of the molten material. The sectioned part of the
305 barrel is either bolted or clamped together. Similarly, the end portion of the barrel is
306 connected to the end-plate die, which is interchangeable depending upon the required shape
307 of the extruded materials [1].

308 With regards to the advantages of HME, it produces a drug in the form of solid dispersion
309 or solution, which could improve solubility of poorly soluble drugs [51]. However, at
310 elevated temperature, there is a high chance of recrystallization of API in the polymer blend
311 as the temperature drops. Using highly viscous molten polymer plasticizer can prevent this
312 problem. Another issue of HME is “Die swell phenomenon” i.e. an increase in the cross-
313 section of the film after ejection from the die depending on the viscoelastic characteristics of
314 polymers. This is due to polymer withstanding high energy kneading and high shear force
315 during extrusion. This problem can be prevented by slowing the speed of screw operation or
316 by gently mixing molten mass for a long time instead of high shear kneading for a short
317 duration [54]. Unlike solvent casting, this method avoids the need of organic solvent; hence,
318 they are proven to be environment friendly [2].

319

320 *6.3. Printing technologies*

321

322 Novel methods such as 3D printing could be used for manufacturing polymeric thin films.
323 It could potentially be a platform for producing the dosage form beneficial to the individual
324 patient. This possibly will resolve the issue of the pharmaceutical industry and pharmacies to
325 meet the future demand of customized medicine [55]. The printing technologies are
326 increasingly gaining popularity because of its flexibility and cost-effectiveness. From the
327 viewpoint of pharmaceutical industry, printing technologies are commonly in practice for
328 identifying or labelling of the pharmaceutical dosage forms, particularly to optimize the
329 product to be readily identified and to prevent counterfeit production. However, this approach
330 has recently been adopted for the drug loading of pharmaceutical dosage forms [3]. The
331 examples include the use of off-the-shelf consumer inkjet printers in which drug-loaded inks
332 are deposited to yield accurately dosed units of pharmaceutical ingredients. In addition, a
333 combination of inkjet and flexographic technologies has been practiced as well [55]. The
334 inkjet printing was used for printing of API on different substrate, whereas the flexographic
335 printing was employed to coat the drug loaded-substrate with a polymeric thin film [56].

336 Anhauser et al. made an attempt to load transdermal patches with drug substances *via*
337 screen printing and pad printing; however, pad printing was limited by the low speed of
338 production. In recent years, inkjet printing has made inroads for preparation of film
339 formulation as a safe and accurate method to produce dosage form of potent drug
340 administered at low dose [57]. Preparation of multiple layer can be done by adding a second
341 printing layer on the top of the first with or without an intermediate base film layer. Further,
342 the printed layer would be shielded by a second base film layer. This will result in modified
343 drug release profiles and protect the ink layer from detachment or mechanical stress during
344 processing like cutting or packaging area [55].

345 Regardless of the various types of printing technique used, all of them contribute to
346 producing a film with more homogeneous distribution and accurate dosage of the drug
347 throughout the films. The dose accuracy and uniform distribution of the drug substances in
348 the films are accounted for several reasons, such as coating mass properties, like viscosity or
349 density, which are inherently influenced by the amount and characteristics of the processed
350 drug substances. With regards to the conventional method of film preparation, it may be very
351 challenging to ensure the same dosage accuracy in the individual units [3]. To summarize,
352 printing a drug on dosage form is the latest intervention for film preparation and it has
353 become a powerful tool to manufacture dosage form with excellent uniformity, speed-ability,
354 and stability. Representing printing technologies that have been used for preparation of
355 polymeric thin films are discussed below.

356

357 *6.3.1. Inkjet printing*

358

359 Inkjet printing is the recently developed technology, which is characterized by its
360 versatility, accuracy, repeatability and relatively inexpensive method that deposits small
361 volumes of solution in films. Inkjet printing is extensively applicable for the preparation of
362 low dose medicines and also offers an opportunity to manufacture personalized medicines
363 [58].

364 Inkjet technology is usually divided into mainly two types: (a) continuous inkjet printing
365 (CIP) and (b) drop on demand (DoD) printing. Both are different in their printing process by
366 which the drops are generated. In the case of CIP, there is a consistent ejection of a liquid
367 through an orifice (nozzle), and it breaks up into a stream of drops under the force of surface
368 tension. For the continuous production of a stream of ink-drops, the individual drop should be
369 'steered' to a particular landing site to produce a printed pattern. This is possible by applying
370 an electric charge on some of the drops that deflect the stream from the main axis under an
371 electrostatic field. On the other hand, ejection of the liquid from the printhead occurs in drop-
372 on-demand printing only when a drop is needed. The production of individual drop takes
373 place rapidly under the response of trigger signal. A DoD printhead consist of multiple
374 nozzles (ranges from 100-1000, even though specialist printhead may have a single nozzle).
375 The drop ejection occurs due to kinetic energy of drop derives from the sources located in the
376 printhead, nearby to each nozzle [59].

377 The uniform distribution and dose accuracy of the drug substance in the film rely upon the
378 density or viscosity of the ink (drug substance solution or suspension), which determine the
379 printability characteristics [3]. Buanz et al. (2011) demonstrated the deposition of low doses
380 of salbutamol sulphate onto commercially available starch-based film using conventional
381 desktop printers [10]. However, inkjet printing is not applicable for high-throughput
382 industrial production, instead using of flexographic printing is regarded more suitable for
383 industrial preparation.

384

385 *6.3.2. Flexographic printing technology (FPT)*

386

387 FPT is a process that transfers active pharmaceutical ingredient into thin films gently *via*
388 contact printing [10]. The flexographic printing is a rotary printing process as depicted in **Fig.**
389 **4**, where ink consisting of drug substance solution and suspension is measured by an anilox

390 roller then are transferred to a printing cylinder that prints the film after unwinding the
391 daughter roll [3]. It is useful for heat sensitive products like proteins and peptides. As the
392 mixing and drying of film formulation are processed before introducing the drug, the
393 problems such as loss of activity of API can be prevented. The production efficiency is also
394 high considering the production rate of 530 oral films per min, hence this process could be
395 expanded to scale-up production [6]. No effect on the mechanical properties of polymeric
396 thin films upon printing drug solutions was witnessed using flexographic printing [57]. In a
397 study, Janßen et al. (2013) found that it was possible to dispense tadalafil and rasagiline
398 mesylate solution onto hydroxypropyl methylcellulose films using flexographic printing. The
399 introduction of hydroxypropyl cellulose appeared to reduce drug crystallization after printing.
400 However, the main drawbacks of flexography are relatively low resolution, high chances of
401 contamination, and the need to prepare a print roller, which is not suitable for large scale
402 production [10].

403

404 **7. Quality issues of thin films**

405

406 For being regarded as an ideal thin film, a film should have adequate flexibility, softness,
407 elasticity, and good physico-chemical stability. Therefore, all these parameters should be
408 considered carefully while developing film to ensure its efficient performance.
409 Characterization of a film is a pre-requisite that may include assessing physical properties
410 such as mechanical strength, hydration, *in vitro* release and other properties. The following
411 section outlines the various critical quality attributes affecting film properties and commonly
412 used *in vitro* methods for film characterization.

413

414 *7.1. Thickness and weight variation*

415

416 The measurement of thickness is necessary as it directly correlates with the amount of drug
417 in the film. In addition, an appropriate thickness is required for the comfortable
418 administration of films. For instance, the ideal thickness of buccal films should be in the
419 range of 50 to 1000 μm [12]. Generally, the thickness of the formed thin films is measured
420 using Vernier caliper, electronic digital micrometer, screw gauge, or scanning electron
421 microscopy (SEM) images [60, 61]. The amount of plasticizer in the formulation is known to
422 increase the film thickness slightly [62]. By inserting m (Batch) – the mass of the whole

423 batch, m (API/film) – the drug amount per film, ρ (Batch) – the density of the formulation, m
 424 (API) – the total drug amount in the batch and A (Film) – the area of one film in **Eq. (1)**, it is
 425 possible to calculate the casting thickness (h). A correction factor f is added due to the shift of
 426 actual value of film thickness compared to the set values. A shift behavior is defined
 427 beforehand over different coating thicknesses [63].

428

$$H (\mu\text{m}) = \frac{m (\text{Batch}) \times m (\text{API/film}) \times 10,000}{\rho (\text{Batch}) \times m (\text{API}) \times A (\text{film})} \quad (1)$$

429

430

431 where, API is active pharmaceutical ingredient, m is mass, ρ is density, and A is area
 432 expressed in g , g/cm^3 , and cm^2 respectively.

433

434 The weight variation is generally carried out to ensure that each film contains the
 435 consistent amount of a drug without significant deviation. It is calculated by weighing the
 436 individual film and the average weights of specified films respectively. The average weight
 437 of film is subtracted from the individual weight of patches. The mean \pm SD values are
 438 calculated for all the formulations. A large variation in weight signifies the inefficiency of the
 439 method applied and high chances are there for non-uniformity in drug content [12].

440

441 7.2. Mechanical and physical properties

442

443 Polymeric films should possess enough tension so that it can be ejected easily from the
 444 pouch, rolled up after casting, and peeled from the release liner, but should not be too flexible
 445 because greater elongation during cutting and packaging might cause variation in film
 446 amount resulting in non-uniformity of API amount per film [49, 64]. Mechanical properties
 447 of films can be defined in terms of Young's modulus, percent elongations, tensile strength
 448 and tear resistance [64, 65]. It has been known that soft and weak polymers exhibit low
 449 tensile strength, low elongation at break and low Young's modulus, whereas, the hard and
 450 tough polymer have a high tensile strength, high elongation at break and high Young's
 451 modulus [11]. Additionally, the mechanical properties of films are affected by the method of
 452 manufacturing and the formulation. In general, some examples of behavior attained from
 453 stress strain curves are showed in **Fig. 5** [6]. The concentration and types of the polymers are

454 largely responsible for producing a film having good mechanical strength and integrity [66].
455 Likewise, the morphological state of the film may alter the mechanical strength, e.g. by
456 crystal growth [64]. Therefore, different factors such as film-forming agent, type of
457 manufacturing process, thickness of film and the type and amount of API in the film have to
458 be considered carefully for controlling the mechanical strength of the film.

459 Blending and cross-linking of two or more polymers are useful methods to improve the
460 mechanical properties of the combined polymeric mix [67]. The film maintains their
461 appearance and integrity after cross-linking, but hardening of the film surface can occur [68].
462 Consistent with this observation, the mechanical properties of PVA-NaCMC films were
463 greater than film composed of PVA or NaCMC alone. The tensile strength of PVA-NaCMC
464 film was found to be 13 to 17 times greater than those of films made of the synthetic polymer
465 N-vinylpyrrolidone [69, 70]. Use of plasticizer may overcome the brittleness and soften the
466 rigidity of the film structure by reducing the intermolecular forces. The most commonly used
467 plasticizer are glycerol, sorbitol, propylene glycol and polyethylene glycol [66, 71]. However,
468 using too much amount of plasticizer can decrease the adhesive strength of films by over-
469 hydrating the film formulations [72]. For example, glycerin intercalates themselves between
470 every individual strand of polymer thereby causing disruption of polymer-polymer
471 interaction. The tertiary structure of the polymers is changed into more flexible and porous
472 type. For this reason, the plasticized polymer deforms at lower tensile strength compared with
473 a polymer without plasticizer [73].

474 In most of the works of literature, most commonly used method for characterizing the
475 mechanical strength of a polymeric film is carried out by using texture analyzer. The system
476 starts measuring force and displacement of the probe when they are in contact with the
477 sample. There is an individual sample holder to aid measurement of small sized film samples
478 (**Fig. 6**). Films are attached by screws between two plates with a cylindrical hole of required
479 diameter. The plate is stabilized to avoid movements using pins, which are placed centrally
480 beneath the punch. The adjustment can be made to move the probe forward according to
481 required working velocity. The measurement starts after the probe is in contact with the
482 sample surface (triggering force). The movement of probe occurs at constant fixed speed until
483 the film detaches. At last, the applied force and displacement (penetration depth) should be
484 recorded along with the room temperature and relative humidity [64]. During the
485 measurement of mechanical strength using texture analyzer, it was found that the contact
486 time, contact force, and the speed of probe withdrawal markedly influence the experimental

487 outcome [74]. The tensile strength is calculated by using several parameters such as folding
488 endurance, percent elongation, elongation at break and Young's modulus.

489

490 7.2.1. Folding endurance

491

492 The flexibility of thin film is important when considering that the films can be
493 administered without breakage. The flexibility of the polymeric thin films can be measured
494 with respect to its folding endurance. The folding endurance is determined by folding the film
495 repeatedly at 180° angle of the plane at the same place until it breaks or folded to 300 times
496 without breaking [75].

497

498 7.2.2. Percent elongation and elongation at break

499

500 Elongation, a kind of deformation, is a simple change in shape that any objects encounter
501 under any applied stress. In other words, when the sample is subjected to tensile stress,
502 deformation of the sample takes place resulting in stretching or elongation of sample [17].
503 Measurement of elongation is generally done to predict the ductility of polymers [65]. Elastic
504 elongation or ultimate elongation of a sample can be measured by using a texture analyzer.
505 Elastic elongation is phenomenon shown by all kinds of elastomers. The percent elongation
506 indicates the stretch ability of material without being broken; whereas, elongation at break
507 means the point until which the film can be stretched when it is torn (or broken) by the
508 applied probe (**Fig. 7**). With the exertion of stress to a sample, strain generates, and the
509 sample elongations will become more predominant as the amount of stress applied increases.
510 However, after reaching to a certain point the sample breaks, this point of breakage is
511 referred as percent elongation break [76]. The formula for percent elongation is given in **Eq.**
512 **(2)** as under:

513

$$\% \text{Elongation} = \left(\frac{\text{Increased length of film}}{\text{Initial length of film}} \right) \times 100 \quad (2)$$

514

515

516 Elongation at break can also be calculated by using following formula as well:

517

518

$$\text{Elongation at break (\%)} = \left(\frac{\sqrt{a'^2 + b^2 + r^2}}{a} - 1 \right) \times 100 \quad (3)$$

519

520

521 where, a is the initial length of the film in the sample holding opening, a' is the length of the
 522 film not punctured by the probe, b is the penetration depth/vertical displacement by the probe
 523 and r is the radius of the probe (**Fig. 7**) [64].

524

525 7.2.3. Young's modulus

526

527 Young's modulus or elastic modulus reflects the stiffness or elasticity of the films. This
 528 indicates resistance to deformation of the films, which can be calculated by plotting the stress
 529 strain curve, where slope indicates the modulus i.e. the greater the slope, greater would be the
 530 tensile modulus. On the other side, the small slope means lesser tensile modulus and
 531 deformation [77]. Simply, a film, exhibiting higher tensile strength and greater Young's
 532 modulus values, is the one which is hard and brittle with small elongation. Texture analyzer
 533 can be used for the measurement of Young's modulus, where slope is obtained from the
 534 stress strain curve. Young's modulus is represented as the ratio of applied stress over strain in
 535 the region of elastic deformation, which can be determined using following formula:

536

$$\text{Young's modulus} = \frac{\text{Slope}}{\text{Film thickness} \times \text{Crosshead speed}} \times 100 \quad (4)$$

537

538

539 A range of crosshead speed can be obtained by changing the speed of the motor of the
 540 texture analyzer [15].

541

542 7.2.4. Tear resistance

543

544 The property of the film to withstand the rupture is known as tear resistance. The
 545 measurement of tear resistance is done by allowing the film to undergo a constant rate of
 546 deformation. The maximum force or stress needed to tear the film is measured in Newton or
 547 pound-force [17]. In a stress strain curve, the area of the plot measures the tear resistance.

548 The relation of an area under the stress strain curve is directly proportional to the toughness
549 of the film i.e. higher area of the plot means the higher toughness of the film and also greater
550 amount of energy that a material can absorb. Therefore, it measures the strength of the
551 material rather than toughness. In fact, a less strong material can be tougher compared with a
552 strong material and no confusion should be created [12].

553

554 7.3. Moisture content

555

556 The amount of moisture in the film could be crucial as it affects the mechanical strength,
557 adhesive properties, and friability of film [78]. Several factors are responsible for elevating
558 water level such as hygroscopic properties of API, polymers, and solvent system used to
559 dissolve the polymeric mixture, and manufacturing techniques. In general, the moisture
560 content of the film is determined by using several methods like Karl Fisher titration or by
561 weighing method. In weighing method, pre-weighed films (initial weight) are heated at a
562 temperature of 100–120 °C until they attain constant weight. Finally, the weight of the final
563 dried sample is taken. The Eq. (5) is used for calculating the amount of moisture content in
564 the film that is expressed as % moisture is given below [12]:

565

$$\text{Moisture content (\%)} = \left[\frac{(\text{Initial weight} - \text{Final dried weight})}{\text{Initial weight}} \right] \times 100 \quad (5)$$

566

567

568 7.4. Swelling

569

570 Swelling properties of films generally observed as the polymers employed for making
571 films are hydrophilic [79]. Swelling of the polymers is known to be the fundamental step
572 required for bioadhesion [80, 81]. In many cases the degree and rate of swelling play a key
573 role in controlling the release of the drug. Hence, these parameters can be considered as the
574 indicator for bioadhesive or mucoadhesive potential and drug release profiles. The testing of
575 swelling is done to measure polymer hydration [82]. Hydrophilic polymers with different
576 structures possess a varying degree of swelling based on the relative resistance of matrix
577 network structure to water molecule movement. For example, a polymer chain having the low
578 ability to form hydrogen bond is unable to form a strong network structure, and water

579 penetration is also difficult to occur. When the number of hydrogen bonds as well as the
 580 strength between the polymers increases, the diffusion of water particles into the hydrated
 581 matrix occurs at a slow rate [83]. This was demonstrated by Panomsuk et al., where he
 582 reported that introduction of mannitol to methylcellulose matrix decreases the swelling index
 583 of the membrane. This may be due to the formation of hydrogen bonding between drugs and
 584 the polymeric matrix [84].

585 Measuring swelling or degree of hydration of the polymeric film plays an important role in
 586 providing key information on the mucoadhesive strength. As we know, the hydration of
 587 polymers are the reasons for relaxation and interpenetration of polymeric chain, however, the
 588 over hydration results in a decrease of mucoadhesion properties due to formation of slippery
 589 mucilage [85]. The swelling properties of films i.e. water absorption capacities are measured
 590 by evaluating the percentage of hydration. For example, the piece of films is weighed (W_1)
 591 and it is subjected to immersion in simulated physiological fluid for a predetermined time.
 592 After the predetermined time, the sample is taken out, wiped off to remove excessive water
 593 on the surface and final weighed is measured (W_2). The calculation is done by using
 594 following formula that is expressed in % [83, 86].

595

$$\text{Hydration (\%)} = \frac{W_2 - W_1}{W_1} \times 100 \quad (6)$$

596

597

598 Furthermore, area swelling ratio (ASR) can be used to determine the swelling property of
 599 the prepared films. As a procedure, the films are placed in a Petri dish and 100 ml quantity of
 600 phosphate buffer (pH=7.4) was poured into it as a swelling fluid. The diameter of a film is
 601 calculated at certain time intervals. The calculation of ASR is based on the **Eq. (7)** [87].

602

$$\text{ASR} = \frac{A_t}{A_0} \quad (7)$$

603

604 where, A_t is area of the film at time t , and A_0 is area of the film at time zero.

605

606 *7.5. Drug release profiles*

607

608 To a great extent, the release kinetics of drugs from the polymer matrix is primarily
609 dependent on the physicochemical properties of the materials used as well as the morphology
610 of the system [36]. Variation in pH or temperature may cause increase or decrease in the
611 erosion or dissolution rates of polymers [88]. Upon contact with biological fluids, the
612 polymeric film starts to swell following polymer chain relaxes resulting in drug diffusion.
613 The release of drug holds a direct relationship with polymer structure; for example, linear
614 amorphous polymers dissolve much faster than cross-linked or partially crystalline polymers
615 [89]. According to several studies, the release of the drug is markedly influenced by erosion
616 of the film. The degradation rate of the film is also dependent on the types of plasticizer [11].
617 For the drug to penetrate the biological membrane, the drug should be released from the
618 delivery systems at an optimum rate. Assessing the drug release from the film is essential as
619 it is the rate-determining step in the process of absorption. The dissolution of drugs and/or
620 films is assessed with the apparatus that are approved for other solid dosage forms [90].

621 In the literature, many authors have done some improvisation on the dissolution apparatus,
622 while others have employed Franz diffusion cells (FDC) for testing the drug release from the
623 polymeric films [12]. A major barrier with respect to film in dissolution testing is the placing
624 of the samples. Several methods have been practiced, where the film are attached on the inner
625 side of the glass vessels or the stirring element using an adhesive tape [91]. Okamoto et al.
626 (2001) conducted a dissolution study of lidocaine film for buccal administration using a JP
627 XIII dissolution apparatus at $37 \pm 0.1^\circ\text{C}$. A film was cut into a circle having an area of 1 cm^2
628 and adhered to a 3 cm diameter weight using double adhesive tape. Then after, the film with
629 weight was placed in a glass vessel filled with 500 ml of artificial saliva so that film dosage
630 form faces upwards as shown in **Fig. 8** [92].

631 632 7.6. Surface morphology

633
634 The morphology of the film should appear homogeneous and continuous to ensure the
635 uniform distribution of drug throughout the polymeric mixture. Self-aggregation might take
636 place during drying because of the intermolecular and convective forces leading to wrinkled
637 surface in films. Additionally, interaction between drug and polymers, and the crystalline
638 nature of the drug may result in the formation of rough surface in the films [93]. Hence,
639 assessing the surface morphology and texture is crucial to assure uniform distribution of
640 drugs without any interaction with the polymers in the film formulation. Various surface
641 characteristics such as surface texture (smooth or rough), thickness, and drug distribution

642 (aggregated or scattered) of the film can be observed using light microscopy, scanning
643 electron microscopy (SEM), transmission electron microscopy (TEM) and related imaging
644 techniques [83]. Amongst all, the scientists have more clung to SEM as a reliable method for
645 examining the surface morphology of the films. The operation is carried out by mounting the
646 films on stubs, sputter coated with gold in an inert environment and subsequently, the
647 photographs are taken at a suitable magnification. This approach can be utilized for close
648 observation of size, shape and the number of pores on the surface of polymeric films. Most
649 recently, there are number of studies on the use of SEM in evaluating the role of chemical
650 composition of the film on the crystallinity, morphology and texture [12].

651

652

653 **8. Packaging of thin films**

654

655 Packaging is crucial to provide mechanical protection as well as to keep the stability of
656 thin film formulations. It acts as a barrier to the moisture, light, and oxygen. A number of
657 choices are available for packaging the polymeric thin films, but not all are effective to
658 preserve the integrity and physical properties of the product. Aluminum foils are most
659 commonly used and considered ideal for film packaging as it prevents the film from moisture
660 and light degradation. Similarly, lidding foil has been employed if tamper proof packaging is
661 needed. Films are subjected to multi-track sealing to achieve an accurate airtight seal between
662 the upper and lower pack foils [17]. The most commonly available sizes of films are 3 x 2
663 cm² and 2 x 2 cm². The packaged films are checked thoroughly before being packed into a
664 secondary packaging container [22]. The packing of manufactured film in foil, paper or
665 plastic pouches is cost-effective, easy to handle, and allows easy formation of the flexible
666 pouch by either vertical or horizontal forming method during product filling [4].

667 Nowadays, the strips are available in both single dose sachets and multiple-unit blisters. A
668 single dose sachet with a name Pocketpaks™ for cool mint Listerine was introduced by
669 Pfizer consumer healthcare. Similarly, a tear notch/slit/cut-off is manufactured to ensure
670 convenience for the consumer to peel-off the pack. This technique is automated and
671 computer-driven process [17]. APR-Labtec launched a patented packaging system with the
672 name Rapid card for the Rapid® films. The rapid card has same size as a credit card and
673 contains three films on each side, which can be removed individually [22].

674

675

676 **9. Routes for the administration of thin films**

677

678 *9.1. Oral route*

679

680 Developing polymeric films have made possible to improve the drug bioavailability and
681 patient adherence to drug therapy *via* the oral route, especially buccal and sublingual route.

682 The anatomical and physiological characteristics of buccal mucosa, such as the existence of
683 smooth muscles with high vascular perfusion, easy accessibility, and bypassing of first pass
684 metabolism make it favorable route for the drug delivery [72]. The oral cavity consists of lips,
685 cheek, tongue, hard palate, soft palate and floor of the mouth [2]. **Fig. 9** demonstrates the
686 common site for administration of films to buccal and sublingual mucosa. Compared with the
687 other mucosa, the buccal and sublingual routes are preferable because it provides better
688 permeability of the drug [94].

689 Squier and co-workers reported that the water penetration across the buccal mucosa to be
690 10 times higher than skin [95]. Similarly, the oral mucosa was found to be 4-4000 times more
691 permeable to a hydrophilic drug than the skin [96]. The sublingual route is targeted for the
692 delivery of drug exhibiting high permeability across the mucosa and is utilized for the
693 treatment of acute disorders. On the other hand, the buccal route is preferred for the treatment
694 of chronic disease, when an extended release of the drug is desired [18]. Direct access to the
695 systemic circulation through the internal jugular vein is possible with buccal drug delivery
696 [36].

697 However, systemic drug delivery in the oral cavity may be extremely challenging due to an
698 unfavorable oral environment and physiological barriers. For achieving a promising
699 therapeutic effect, the drug must be released from the formulation to the delivery site (e.g.
700 sublingual or buccal region) and should penetrate the oral mucosa to reach the systemic
701 circulation. The existence of several environmental related factors such as fluid volume, pH,
702 enzyme activity and the permeability of oral mucosa determines the fate of drug absorption in
703 the oral mucosa. On the other side, the amount of secretion of saliva impedes the residence
704 time of drug at the delivery site due to washing out of the drug. Similarly, the swallowing of
705 drugs might occur before the absorption of the drug through the oral mucosa [2, 97]. Hence,
706 while developing the oral formulation like polymeric films, all the point should be taken into
707 account for obtaining higher therapeutic bioavailability as well as the patient adherence to the
708 dosage form.

709 Films containing the polymeric blend would be an ideal platform for the delivery of drugs
710 in the oral cavity because of its comfort and flexibility [98]. Over the last decade, there has
711 been an enormous rise in the development of buccal films as an alternative drug delivery for
712 various classes such as anti-inflammatory, analgesics, anesthetic drugs and proteins and
713 peptides. Of recent, mucoadhesive films have been used as a delivery platform for
714 transmucosal buccal delivery of Biopharmaceutics Classification System (BCS) Class II
715 drugs particularly targeting the opioid analgesics like fentanyl citrate, which is available with
716 a trademark name such as Onsolis®/Breakyl® for treating immense pain [26]. Similarly, the
717 mucoadhesive film remains attached to the buccal area without showing any erratic
718 absorption profile resulting in less inter and intra-individual variability [72]. Oral thin films
719 (OTFs) are comparable to the disintegrating system, which is soaked in saliva and stick to the
720 site of application. The rate of disintegration is rapid allowing the drug to release and
721 followed by the oromucosal absorption. Many drugs that undergo degradation in the GI tract
722 are being administered employing this route [99].

723 In context to the commercially marketed product of the oral thin film, the nutraceuticals
724 and over-the-counter drugs were among the first to be introduced in the market, and included
725 the incorporated active such as vitamins, herbal and non-herbal extracts. In 2001, Pfizer
726 introduced a thin film product of Listerine pocketpaks® developed as mouth freshener. The
727 company Bio-film has been putting an endeavor to develop oral thin films. Not only the
728 pharmaceuticals but they are also using nutraceuticals such as vitamins, aphrodisiac, energy
729 boosters, and appetite suppressor that targets a specific population of the certain age group.
730 The energy booster consists of various compounds such as caffeine, guarana, and green tea
731 extract to maintain the energy levels [17]. A number of companies have been attempting to
732 develop a drug delivery platform based on polymeric films. Most of them have already
733 succeeded in obtaining a film with rapid release along with better therapeutic outcomes [2].
734 The companies with their technology platform based on polymeric film are listed in the
735 **Table 2.**

736

737 *9.2. Ocular route*

738

739 More than 90% of the marketed ocular formulation are in the form of solutions or
740 suspension; however, this conventional dosage form lacks to achieve promising therapeutic
741 success [100]. The frequent instillation of eye drops is needed to elicit a therapeutic response.
742 This usually leads to patient non-compliance and pulsed administration. Furthermore, the

743 topically applied drugs to the eye generally enter the systemic circulation *via* the nasolacrimal
744 duct system that possibly cause side effects and systemic toxicity as well [101]. With the aim
745 of enhancing the ocular bioavailability and overcoming the ocular drug delivery barriers, the
746 development of ophthalmic film becomes popular these days [84]. The ophthalmic films
747 result in the reduction of dose frequency, less systemic side effects and better therapeutic
748 outcomes. Therefore, ophthalmic films could open the exciting opportunities as a delivery
749 platform of therapeutics to replace the traditional dosage forms for achieving high therapeutic
750 success and patient adherence. So far, the list of drugs formulated in ophthalmic films is
751 presented below in **Table. 3**.

752 The flow of tear across the outer surface of the cornea is continuous, which impedes the
753 drug diffusion leading in low bioavailability (1-7%) of drugs [109]. Generally, the drug with
754 higher lipophilicity encounters many problems as it cannot be dissolved in the aqueous
755 medium of the eye. Since the drug causes discomfort in the eye, it induces blinking and
756 therefore, causing washing out of the significant amount of drug. Therefore, the success of
757 the effective development of films to be delivered to the eye relies on the comprehensive
758 knowledge of the drug, the constraints to ocular drug delivery, and the excipients used.
759 Hence, all these factors should be considered during the formulation of ocular films.

760

761 9.3. *Transdermal route*

762

763 Drug-loaded transdermal films are the alternative to replace the existing transdermal
764 dosage form. Numerous sustained or controlled delivery systems have been devised, where a
765 drug is either dissolved or dispersed in the films [71]. The film-forming system has been
766 practiced for the transdermal delivery of steroidal hormones, analgesics, local anesthesia and
767 anti-emetic for systemic effects [110, 111, 112].

768 Only a small number of drugs are being designed for the transdermal delivery of films as
769 several factors affect the bioavailability of drug such as molecular size, polarity, pH of the
770 drug, state of the skin hydration, subcutaneous reservoir of drug and drug metabolism by skin
771 flora [113]. Similarly, the hydration of skin is crucial for increasing drug absorption, which is
772 possible by using humectant in the film formulation. The physiological factors such as
773 regional skin site, nature of stratum corneum, the thickness of skin, and density of
774 appendages also influence the overall outcome of the therapeutic effects of the drug [114].

775 The thin film may possess better therapeutic efficacy and patient acceptance compared to
776 the common transdermal dosage forms such as patches or gels [115]. Due to occlusive

777 properties of transdermal patches, it prevents the permeation of water vapour from the skin
778 surface and causes severe pain at the time of peeling. However, polymeric thin films could be
779 a highly promising alternative for transdermal drug delivery because of the ease of
780 application, flexibility and better cosmetic appearance [29].

781

782 **10. Future scope of development and conclusion**

783

784 The formulation of a drug into various films has been popular in recent years. Several
785 undesirable drawbacks associated with conventional dosage forms such as inconvenience of
786 administration, lower bioavailability and patient non-compliance have pushed to the
787 development of novel polymeric thin films as a drug delivery platform. This drug delivery
788 platform is being under surveillance from both start-up and established pharmaceutical
789 companies. The companies strive to design a wide range of thin films for oral, buccal,
790 sublingual, ocular and transdermal routes. Therefore, as an alternative to conventional dosage
791 forms polymeric thin films are expected to stand out as a dosage form to overcome the
792 limitations posed by existing dosage forms. The film dosage form encounters several
793 challenges during the phases of formulation development and manufacture. Such issues
794 should be addressed to optimize the overall formulation even after transferring to large scale
795 manufacturing. The future looks very promising for the film technology in the time to come
796 as new technologies are rapidly introduced to prepare thin films.

797

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803

804 **Conflict of Interest**

805 The authors declare no conflict of interest.

806

807 **References**

808

809 [1] Maniruzzaman M, Boateng JS, Snowden MJ, Douroumis D. A review of hot-melt
810 extrusion: process technology to pharmaceutical products. *ISRN Pharm* 2012;2012:1–
811 9.

812 [2] Patel VF, Liu F, Brown MB. Advances in oral transmucosal drug delivery. *J Control*
813 *Release* 2011;153:106–116.

814 [3] Borges AF, Silva C, Coelho JFJ, et al. Oral films: Current status and future
815 perspectives. *J Control Release* 2015;206:1–19.

816 [4] Sharma D, Kaur D, Verma S, et al. Fast Dissolving Oral Films Technology : A Recent
817 Trend For An Innovative Oral Drug Delivery System. *Int J drug Deliv* 2015;7:60–75.

818 [5] Kang-Mieler JJ, Osswald CR, Mieler WF. Advances in ocular drug delivery: emphasis
819 on the posterior segment. *Expert Opin Drug Deliv* 2014;11:1–14.

820 [6] Castro PM, Fonte P, Sousa F, et al. Oral films as breakthrough tools for oral delivery
821 of proteins/peptides. *J Control Release* 2015;211:63–73.

822 [7] Barbu E, Verestiuc L, Nevell TG, Tsibouklis J. Polymeric materials for ophthalmic
823 drug delivery: trends and perspectives. *J Mater Chem*. 2006;16:3439-3443.

824 [8] Achouri D, Alhanout K, Piccerelle P, et al. Recent advances in ocular drug delivery.
825 *Drug Dev Ind Pharm* 2013;39:1599–1617.

826 [9] Hearnden V, Sankar V, Hull K, Juras DV, Greenberg M, Kerr AR, et al. New
827 developments and opportunities in oral mucosal drug delivery for local and systemic
828 disease. *Adv Drug Deliv Rev*. 2012;64:16–28.

829 [10] Buanz ABM, Belaunde CC, Soutari N, et al. Ink-jet printing versus solvent casting to
830 prepare oral films: Effect on mechanical properties and physical stability. *Int J Pharm*
831 2015;494:611–618.

832 [11] Morales JO, McConville JT. Manufacture and characterization of mucoadhesive
833 buccal films. *Eur J Pharm Biopharm* 2011;77:187–199.

- 834 [12] Nair AB, Kumria R, Harsha S, et al. In vitro techniques to evaluate buccal films. J
835 Control Release 2013;166:10–21.
- 836 [13] Ng YC, Yang Z, McAuley WJ, et al. Stabilisation of amorphous drugs under high
837 humidity using pharmaceutical thin films. Eur J Pharm Biopharm 2013;84:555–565.
- 838 [14] Kumari A, Sharma P, Garg V, et al. Ocular inserts - Advancement in therapy of eye
839 diseases. J Adv Pharm Technol Res 2010;1:291-296.
- 840 [15] Irfan M, Rabel S, Bukhtar Q, Qadir MI, Jabeen F, Khan A. Orally disintegrating films:
841 A modern expansion in drug delivery system. Saudi Pharm J. King Saud University
842 2015;2015:1-10.
- 843 [16] Patel A, Cholkar K, Agrahari V, et al. Ocular drug delivery systems: An overview.
844 World J Pharmacol 2015;2:47–64.
- 845 [17] Dixit RP, Puthli SP. Oral strip technology: Overview and future potential. J Control
846 Release 2009;139:94–107.
- 847 [18] Hao J, Heng PWS. Buccal delivery systems. Drug Dev Ind Pharm [Internet].
848 2003;29:821–32.
- 849 [19] Rathore KS, Nema RK, Sisodia SS. Timolol maleate a gold standard drug in glaucoma
850 used as ocular films and inserts: An overview. Int J Pharm Sci Rev Res. 2010;3:23–9.
- 851 [20] Juluru N. Fast Dissolving Oral Films : A Novel Drug Delivery System. Int J Pharm Sci
852 Rev Res 2013;2:108–112.
- 853 [21] Siddiqui MDN, Garg G, Sharma PK. A Short Review on “A Novel Approach in Oral
854 Fast Dissolving Drug Delivery System and Their Patents”. Adv Biol Res 2011;5:291–
855 303.
- 856 [22] Amin PM, Gangurde AB, Alai P V. Oral Film Technology : Challenges and Future
857 Scope for Pharmaceutical Industry. Int J Pharm Pharm Res 2015;3:183–203.
- 858 [23] Vibhooti P, Preeti K. Wafers Technology–A newer approach to smart drug delivery
859 system. IJRPB 2013;1:428–439.
- 860 [24] Hoffmann EM, Breitenbach A, Breitreutz J. Advances in orodispersible films for

- 861 drug delivery. *Expert Opin Drug Deliv* 2011;8:299–316.
- 862 [25] Prabhu SC, Parsekar SD, Shetty A, et al. Review Article A Review on Fast Dissolving
863 Sublingual Films for Systemic Drug Delivery 2014;3:501–511.
- 864 [26] Russo E, Selmin F, Baldassari S, et al. A focus on mucoadhesive polymers and their
865 application in buccal dosage forms. *J Drug Deliv Sci Technol* 2015;32:113-125.
- 866 [27] Wening K, Breitzkreutz J. Oral drug delivery in personalized medicine: Unmet needs
867 and novel approaches. *Int J Pharm* 2011;404:1–9.
- 868 [28] Sultana Y, Jain R, Aqil M, Ali A. Review of ocular drug delivery. *Curr Drug Deliv.*
869 2006;3:207–217.
- 870 [29] Guo R, Du X, Zhang R, et al. Bioadhesive film formed from a novel organic-inorganic
871 hybrid gel for transdermal drug delivery system. *Eur J Pharm Biopharm* 2011;79:574-
872 583.
- 873 [30] Boddupalli B, Mohammed Z, Nath R, et al. Mucoadhesive drug delivery system: An
874 overview. *J Adv Pharm Technol Res.* 2010;1:381-387.
- 875 [31] Rawas-Qalaji M, Williams CA. Advances in ocular drug delivery. *Curr Eye Res*
876 2012;37:345–356.
- 877 [32] Jadhav YG, Galgatte UC, Chaudhari PD. Challenges in formulation development of
878 fast dissolving oral films. *Indo Am J Pharm Res* 2013;3:1746–1751.
- 879 [33] Perumal VA, Govender T, Lutchman D, et al. Investigating a New Approach to Film
880 Casting for Enhanced Drug Content Uniformity in Polymeric Films. *Drug Dev Ind*
881 *Pharm.* 2008;34:1036–1047.
- 882 [34] Renukuntla J, Vadlapudi AD, Patel A, et al. Approaches for enhancing oral
883 bioavailability of peptides and proteins. *Int J Pharm* 2013;447:75–93.
- 884 [35] Khairnar GA, Sayyad FJ. Development of buccal drug delivery system based on
885 mucoadhesive polymers. *Int J PharmTech Res* 2010;2:719–735.
- 886 [36] Sudhakar Y, Kuotsu K, Bandyopadhyay AK. Buccal bioadhesive drug delivery - A
887 promising option for orally less efficient drugs. *J Control Release* 2006;114:15–40.

- 888 [37] Dingel A, Nagarsenker M. Formulation and Evaluation of Fast Dissolving Films for
889 Delivery of Triclosan to the Oral Cavity. *AAPS PharmSciTech* 2008;9:349–356.
- 890 [38] Cilurzo F, Cupone IE, Minghetti P, et al. Fast dissolving films made of maltodextrins.
891 *Eur J Pharm Biopharm* 2008;70:895–900.
- 892 [39] Chaudhary H, Gauri S, Rathee P, Kumar V. Development and optimization of fast
893 dissolving oro-dispersible films of granisetron HCl using Box – Behnken statistical
894 design. *Bull Fac Pharmacy, Cairo Univ* 2013;51:193–201.
- 895 [40] Landová H, Vetchý V, Gajdziok J. Evaluation of the influence of formulation and
896 process variables on mechanical properties of oral mucoadhesive films using
897 multivariate data analysis. *Biomed Res Int.* 2014;2014:1–9.
- 898 [41] Smart JD, Kellaway IW, Worthington HEC. An in-vitro investigation of mucosa-
899 adhesive materials for use in controlled drug delivery. *J Pharm Pharmacol*
900 1984;36:295–299.
- 901 [42] Ludwig A. The use of mucoadhesive polymers in ocular drug delivery. *Adv Drug*
902 *Deliv Rev* 2005;57:1595–1639.
- 903 [43] Fefelova NA, Nurkeeva ZS, Mun GA, et al. Mucoadhesive interactions of amphiphilic
904 cationic copolymers based on [2-(methacryloyloxy)ethyl]trimethylammonium
905 chloride. *Int J Pharm* 2007;339:25–32.
- 906 [44] Cook MT, Khutoryanskiy V V. Mucoadhesion and mucosa-mimetic materials—A
907 mini-review. *Int J Pharm* 2015;495:991–998.
- 908 [45] Juliano C, Cossu M, Pigozzi P, et al. In Vitro Characterization and Preliminary In Vivo
909 Evaluation of Buccal Polymeric Films Containing Chlorhexidine. *AAPS*
910 *PharmSciTech.* 2008;9:1153–1158.
- 911 [46] Kunte S, Tandale P. Fast dissolving strips: A novel approach for the delivery of
912 verapamil. *J Pharm bioallied Sci* 2010;2:325–328.
- 913 [47] El-Setouhy DA, Shakwy N, El-Malak ABD. Formulation of a novel tianeptine sodium
914 orodispersible film. *AAPS PharmSciTech* 2010;11:1018–1025.

- 915 [48] Low AQJ, Parmentier J, Khong YM, et al. Effect of type and ratio of solubilising
916 polymer on characteristics of hot-melt extruded orodispersible films. *Int J Pharm*
917 2013;455:138–147.
- 918 [49] Preis M, Woertz C, Kleinebudde P, et al. Oromucosal film preparations: classification
919 and characterization methods. *Expert Opin Drug Deliv* 2013;10:1–15.
- 920 [50] Verma S, Kumar N, Sharma PK. Buccal Film : An Advance Technology for Oral Drug
921 Delivery. *Advan. Biol. Res* 2014;8:260–267.
- 922 [51] Repka MA, Gutta K, Prodduturi S, et al. Characterization of cellulosic hot-melt
923 extruded films containing lidocaine. *Eur J Pharm Biopharm* 2005;59:189–196.
- 924 [52] Crowley MM, Zhang F. Pharmaceutical Applications of Hot-Melt Extrusion: Part I.
925 *Drug Dev Ind Pharm* 2007;33:909–926.
- 926 [53] Chokshi R, Zia H. Hot-Melt Extrusion technique: A Review. *Iran J Pharm Res*
927 2004;3:3–16.
- 928 [54] Jani R, Patel D. Hot melt extrusion: An industrially feasible approach for casting
929 orodispersible film. *Asian J Pharm Sci* 2014;10:292–305.
- 930 [55] Preis M, Breitzkreutz J, Sandler N. Perspective: Concepts of printing technologies for
931 oral film formulations. *Int J Pharm* 2015;494:578–584.
- 932 [56] Genina N, Fors D, Vakili H, et al. Tailoring controlled-release oral dosage forms by
933 combining inkjet and flexographic printing techniques. *Eur J Pharm Sci* 2012;47:615–
934 623.
- 935 [57] Janßen EM, Schliephacke R, Breitenbach A, et al. Drug-printing by flexographic
936 printing technology - A new manufacturing process for orodispersible films. *Int J*
937 *Pharm* 2013;441:818–825.
- 938 [58] Alomari M, Mohamed FH, Basit AW, et al. Personalised dosing: Printing a dose of
939 one's own medicine. *Int J Pharm* 2015;494:568–577.
- 940 [59] Daly R, Harrington TS, Martin GD, et al. Inkjet printing for pharmaceuticals - A review
941 of research and manufacturing. *Int J Pharm* 2015;494:554–567.

- 942 [60] Ali J, Arora S, Ahuja A, Babbar AK, et al. Formulation and development of floating
943 capsules of celecoxib: in vitro and in vivo evaluation. *AAPS PharmSciTech*
944 2007;8:E1-E8.
- 945 [61] Prabhushankar GL, Gopalkrishna B, Manjunatha KM, et al. Formulation and
946 evaluation of levofloxacin dental films for periodontitis. *Int J Pharm Pharm Sci*
947 2010;2:162–168.
- 948 [62] Cao N, Yang X, Fu Y. Effects of various plasticizers on mechanical and water vapor
949 barrier properties of gelatin films. *Food Hydrocoll* 2009;23:729–735.
- 950 [63] Preis M, Pein M, Breitzkreutz J. Development of a taste-masked orodispersible film
951 containing dimenhydrinate. *Pharmaceutics* 2012;4:551–562.
- 952 [64] Preis M, Knop K, Breitzkreutz J. Mechanical strength test for orodispersible and buccal
953 films. *Int J Pharm* 2014;461:22–29.
- 954 [65] Heng PWS, Chan LW, Ong KT. Influence of storage conditions and type of
955 plasticizers on ethylcellulose and acrylate films from aqueous dispersions. *J Pharm*
956 *Pharm Sci.* 2003;6:334–344.
- 957 [66] Liew KB Tan YTF, Peh KK. Effect of polymer, plasticizer and filler on orally
958 disintegrating film. *Drug Dev Ind Pharm* 2014;40:110–119.
- 959 [67] Dong Z, Wang Q, Du Y. Alginate/gelatin blend films and their properties for drug
960 controlled release. *J Memb Sci* 2006;280:37–44.
- 961 [68] Mathurm M, Gilhotra RM. Glycerogelatin-based ocular inserts of aceclofenac:
962 physicochemical, drug release studies and efficacy against prostaglandin E₂-induced
963 ocular inflammation. *Drug Deliv* 2011;18:54–64.
- 964 [69] Jain D, Carvalho E, Banerjee R. Biodegradable hybrid polymeric membranes for
965 ocular drug delivery. *Acta Materialia Inc* 2010;6:1370–1379.
- 966 [70] Karolewicz B. A review of polymers as multifunctional excipients in drug dosage form
967 technology. *Saudi Pharm J.* 2015;2015:1–12.
- 968 [71] Nesseem DI, Eid SF, El-Houseny SS. Development of novel transdermal self-adhesive

- 969 films for tenoxicam, an anti-inflammatory drug. *Life Sci. Elsevier Inc.*; 2011;89:430–
970 438.
- 971 [72] Kaur G, Singh D, Brar V. Bioadhesive okra polymer based buccal patches as platform
972 for controlled drug delivery. *Int J Biol Macromol* 2014;70:408–419.
- 973 [73] Laohakunjit N, Noomhorm A. Effect of plasticizers on mechanical and barrier
974 properties of rice starch film. *Starch/Staerke* 2004;56:348–356.
- 975 [74] Wong CF, Yuen KH, Peh KK. An in-vitro method for buccal adhesion studies:
976 Importance of instrument variables. *Int J Pharm* 1999;180:47–57.
- 977 [75] Mukherjee D, Bharath S. Design and Characterization of Double Layered
978 Mucoadhesive System Containing Bisphosphonate Derivative. *ISRN Pharm*
979 2013;2013:1–10.
- 980 [76] Fulzele SV., Satturwar PM, Dorle AK. Polymerized rosin: Novel film forming
981 polymer for drug delivery. *Int J Pharm* 2002;249:175–184.
- 982 [77] Yan W, Wen S, Liu J, et al. Determination of reduced Young's modulus of thin films
983 using indentation test. *Acta Metall Sin* 2009;22:468–480.
- 984 [78] Gorle AP, Gattani SG. Development and evaluation of ocular drug delivery system.
985 *Pharm Dev Technol* 2010;15:46–52.
- 986 [79] Peh KK, Wong CF. Polymeric films as vehicle for buccal delivery: swelling,
987 mechanical, and bioadhesive properties. *J Pharm Pharm Sci* 1999;2:53–61.
- 988 [80] Roy SK, Prabhakar B. Bioadhesive polymeric platforms for transmucosal drug
989 delivery systems - A review. *Trop J Pharm Res.* 2010;9:91–104.
- 990 [81] Kumria R, Nair AB, Goomber G, et al. Buccal films of prednisolone with enhanced
991 bioavailability. *Drug Deliv* 2014;23:471–478.
- 992 [82] Aburahma MH, Mahmoud AA. Biodegradable Ocular Inserts for Sustained Delivery
993 of Brimonidine Tartarate: Preparation and In Vitro/In Vivo Evaluation. *AAPS*
994 *PharmSciTech* 2011;12:1335–1347.
- 995 [83] Baranowski P, Karolewicz B, Gajda M, et al. Ophthalmic drug dosage forms:

- 996 Characterisation and research methods. *Sci World J* 2014;2014:1-14.
- 997 [84] Franca JR, Foureaux G, Fuscaldi LL, et al. Bimatoprost-loaded ocular inserts as
998 sustained release drug delivery systems for glaucoma treatment: In Vitro and in Vivo
999 evaluation. *PLoS One* 2014;9:1–11.
- 1000 [85] Mortazavi SA, Smart JD. An investigation into the role of water movement and mucus
1001 gel dehydration in mucoadhesion. *J Control Release* 1993;25:197–203.
- 1002 [86] Eroğlu H, Sargon MF, Öner L. Chitosan Formulations for Steroid Delivery: Effect of
1003 Formulation Variables on In Vitro Characteristics. *Drug Dev Ind Pharm* 2007;33:265–
1004 271.
- 1005 [87] Attama AA, Akpa PA, Onugwu LE, et al. Novel buccoadhesive delivery system of
1006 hydrochlorothiazide formulated with ethyl cellulose- hydroxypropyl methylcellulose
1007 interpolymer complex. *Sci Res Essay*. 2008;3:343–347.
- 1008 [88] Taylor MJ, Tanna S, Sahota T. In vivo study of a polymeric glucose-sensitive insulin
1009 delivery system using a rat model. *J Pharm Sci* 2010;99:4215–4227.
- 1010 [89] Tangri P, Khurana S. Basics of Ocular Drug Delivery Systems. *Int J Res Pharm*
1011 *Biomed Sci* 2011;2:1541–1552
- 1012 [90] Adrover A, Pedacchia A, Petralito S, et al. In vitro dissolution testing of oral thin
1013 films: A comparison between USP 1, USP 2 apparatuses and a new millifluidic flow-
1014 through device. *Chem Eng Res Des* 2015;95:173–178.
- 1015 [91] Hoffmann EM, Breitenbach A, Breitzkreutz J. Advances in orodispersible films for
1016 drug delivery. *Expert Opin Drug Deliv* 2011;8:299–316.
- 1017 [92] Okamoto H, Taguchi H, Iida K, et al. Development of polymer film dosage forms of
1018 lidocaine for buccal administration I. Penetration rate and release rate. *J Control*
1019 *Release* 2001;77:253-260.
- 1020 [93] Hermans K, Van Den Plas D, Kerimova S, et al. Development and characterization of
1021 mucoadhesive chitosan films for ophthalmic delivery of cyclosporine A. *Int J Pharm*
1022 2014;472:10–19.

- 1023 [94] Gandhi RB, Robinson JR. Oral cavity as a site for bioadhesive drug delivery. *Adv*
1024 *Drug Deliv Rev* 1994;13:43–74.
- 1025 [95] Lesch CA, Squier CA, Cruchley A, et al. The Permeability of Human Oral Mucosa and
1026 Skin to Water. *J Dent Res* 1989;68:1345–1349.
- 1027 [96] Galey WR, Lonsdale HK, Nacht S. The In Vitro Permeability Of Skin And Buccal
1028 Mucosa To Selected Drugs And Tritiated Water. *J Invest Dermatol* 1976;67:713–717.
- 1029 [97] Lam JKW, Xu Y, Worsley A, et al. Oral transmucosal drug delivery for pediatric use.
1030 *Adv Drug Deliv Rev* 2014;73:50–62.
- 1031 [98] Perumal VA, Lutchman D, Mackraj I, et al. Formulation of monolayered films with
1032 drug and polymers of opposing solubilities. *Int J Pharm* 2008;358:184–191.
- 1033 [99] Haque SE, Sheela A. Development of polymer-bound fast-dissolving metformin
1034 buccal film with disintegrants. *Int J Nanomedicine* 2015;10:199–205.
- 1035 [100] de la Fuente M, Raviña M, Paolicelli P, et al. Chitosan-based nanostructures: A
1036 delivery platform for ocular therapeutics. *Adv Drug Deliv Rev* 2010;62:100–117.
- 1037 [101] Mahajan HS, Deshmukh SR. Development and evaluation of gel-forming ocular films
1038 based on xyloglucan. *Carbohydr Polym* 2015;122:243–247.
- 1039 [102] Al-snafi AE. International Journal of Pharmaceuticals and Health care Research. *Int J*
1040 *Pharm Heal care Res* 2014;02:162–172.
- 1041 [103] Rathore KS, Nema RK, Sisodia SS. Preparation and characterization of timolol
1042 maleate ocular films. *Int J PharmTech Res.* 2010;2:1995–2000.
- 1043 [104] de Oliveira Fulgêncio G, Viana FA, Silva RO, et al. Mucoadhesive chitosan films as a
1044 potential ocular delivery system for ofloxacin: Preliminary in vitro studies. *Vet*
1045 *Ophthalmol.* 2014;17:150–155.
- 1046 [105] Tandale YN, Wagh VD. Formulation and Evaluation of Dorzolamide hydrochloride
1047 Polymeric Film. *Int J PharmTech Res.* 2011;3:1817–1824.
- 1048 [106] Dubey BK, Bhowmick M, Joshi A, et al. Design and characterization of sustained
1049 release levofloxacin ocular inserts. *Int J Biomed Adv* 2012;02:453-466.

- 1050 [107] Sharma S, Parvez N, Sharma PK. Formulation and Evaluation of Naphazoline HCl
1051 Ocular Insert. *Glob J Pharmacol* 2015;9:97–101.
- 1052 [108] Rajasekaran A, Sivakumar V, Karthika K, et al. Design and Evaluation of Polymeric
1053 Controlled Release. *Eng Technol*. 2010;6:108–115.
- 1054 [109] Kim YC, Chiang B, Wu X, et al. Ocular delivery of macromolecules. *J Control*
1055 *Release* 2014;190:172–181.
- 1056 [110] Padula C, Nicoli S, Colombo P, et al. Single-layer transdermal film containing
1057 lidocaine: Modulation of drug release. *Eur J Pharm Biopharm* 2007;66:422–428.
- 1058 [111] Ammar HO, Ghorab M, Mahmoud AA, et al. Rapid pain relief using transdermal film
1059 forming polymeric solution of ketorolac. *Pharm Dev Technol* 2013;18:1005–1016.
- 1060 [112] Aktar B, Erdal MS, Sagirli O, et al. Optimization of biopolymer based transdermal
1061 films of metoclopramide as an alternative delivery approach. *Polymers* 2014;6:1350-
1062 1365.
- 1063 [113] Tanner T, Marks R. Delivering drugs by the transdermal route: Review and comment.
1064 *Ski Res Technol* 2008;14:249–260.
- 1065 [114] Mali ADM, Bathe R, Patil M. An updated review on transdermal drug delivery
1066 systems. *Int J Adv Sci Res* 2015;1:244–254.
- 1067 [115] Schroeder IZ, Franke P, Schaefer UF, et al. Delivery of ethinylestradiol from film
1068 forming polymeric solutions across human epidermis in vitro and in vivo in pigs. *J*
1069 *Control Release* 2007;118:196–203.
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1072 **Figure legends**

1073 **Fig. 1.** Solvent casting method for film preparation with quality control parameters in each
1074 step

1075 **Fig. 2.** Commercial manufacturing of film based on solvent-casting (reproduced from Amin
1076 et al., 2015 [22])

1077 **Fig. 3.** Holt-melt extrusion system for the preparation of films (reproduced from Amin et al.,
1078 2015 [22])

1079 **Fig. 4.** Schematic overview of flexography technology for the preparation of films
1080 (reproduced from Janßen et al., 2013 [57])

1081 **Fig. 5.** Examples of stress-strain curves obtained from polymeric thin films (reproduced from
1082 Morales & McConville, 2011 [11])

1083 **Fig. 6.** Experimental setup (left) and sample holder for the film preparation (right), where r_s
1084 indicates radius of samples, and r_p indicates radius of probe. Geometry of cylindrical probes
1085 A and B and spherical probe C is shown on the right bottom (reproduced from Preis et al.,
1086 2014 [64]).

1087 **Fig. 7.** Determination of percent elongation of thin films using a texture analyzer, where a =
1088 initial length of the film in the sample holder opening, a' = initial length - radius of probe, b =
1089 displacement of the probe, $c' + r$ = length after strain, c' = length of a' after strain, r = radius
1090 of the probe [64]

1091 **Fig. 8.** Schematic illustration of the apparatus used for dissolution studies of films. The film
1092 dosage form (1 cm^2) was attached to a 3 cm diameter weight using double adhesive tape
1093 (reproduced from Okamoto et al., 2001 [92]).

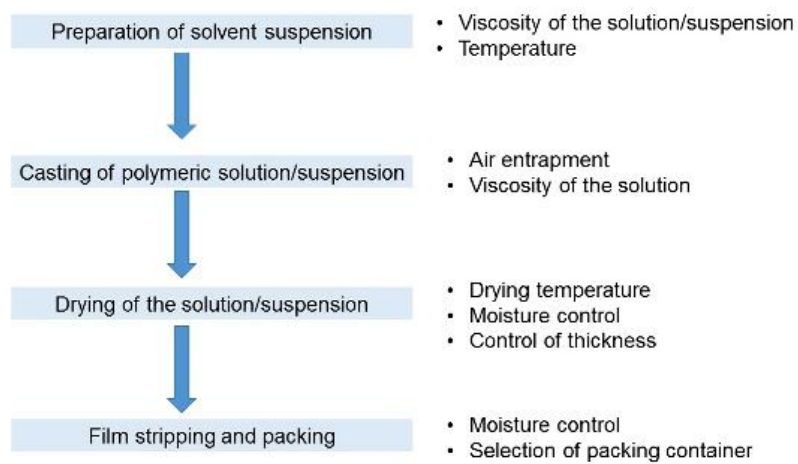
1094 **Fig. 9.** Demonstration of common site for application of film in buccal and sublingual
1095 mucosa (reproduced from Lam et al., 2014 [97])

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1097

1098 **Fig. 1**

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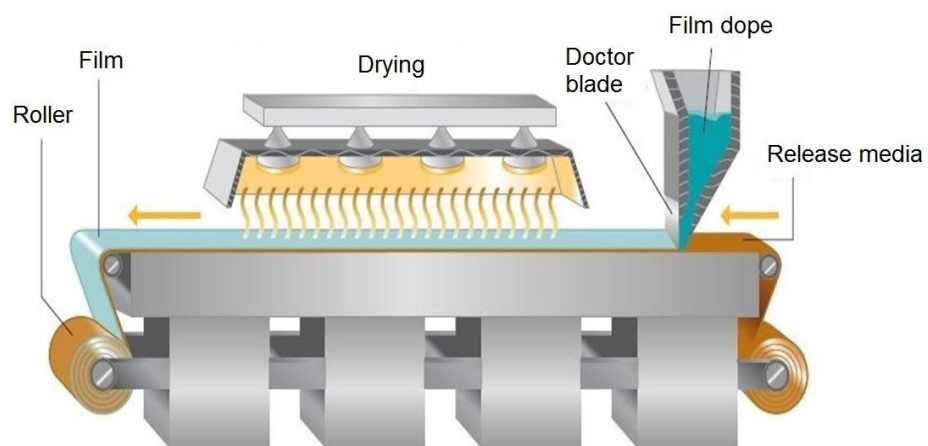
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1102 **Fig. 2**

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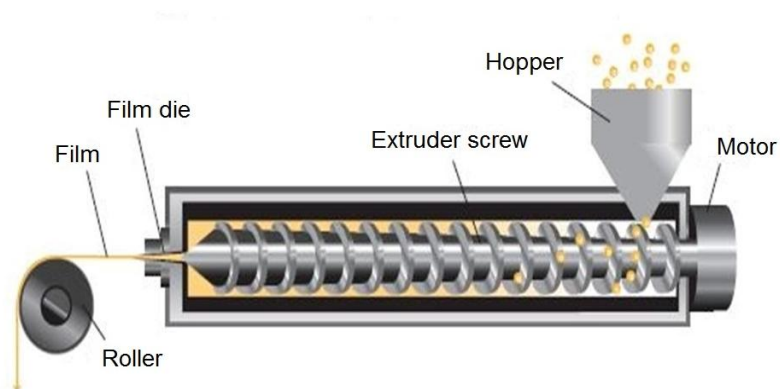
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1107 **Fig. 3**

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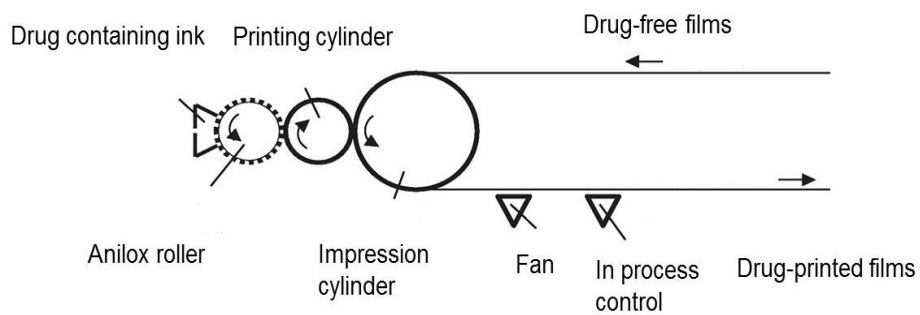


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1111 **Fig. 4**

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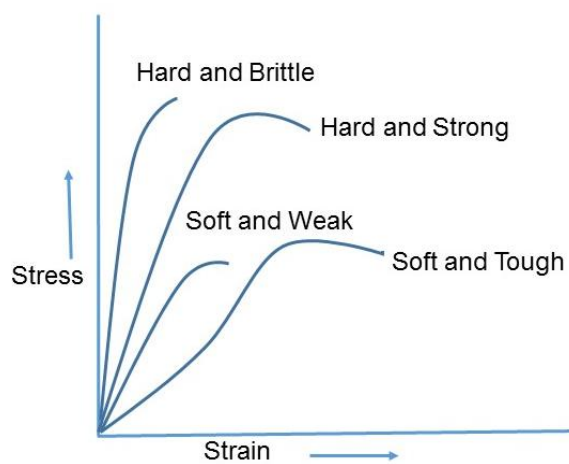
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1116 Fig. 5

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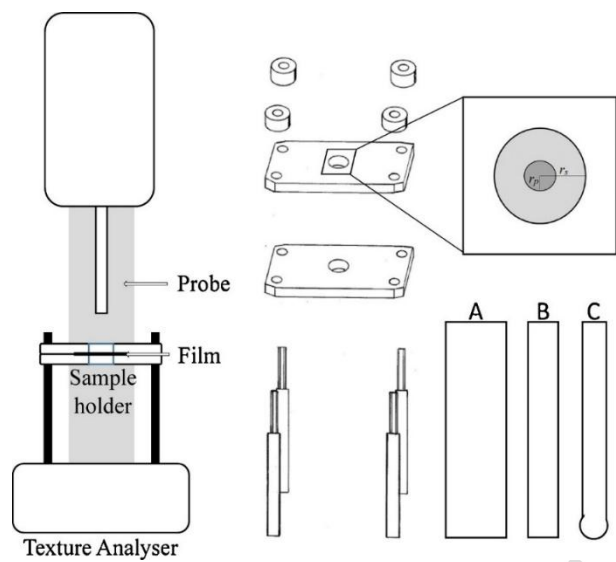
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1121 **Fig. 6**

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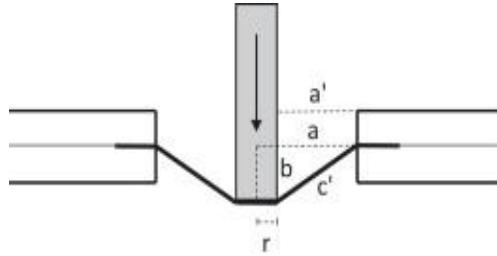


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1125 **Fig. 7**

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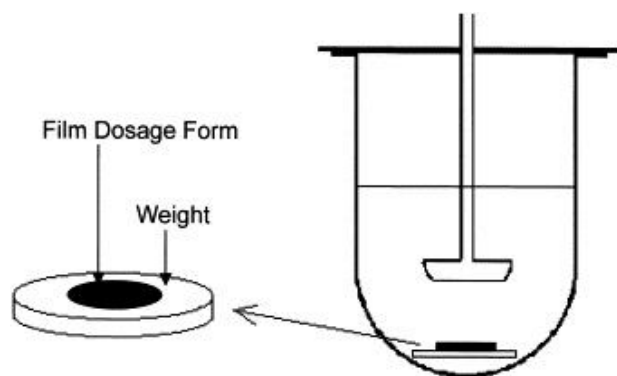
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1130 Fig. 8

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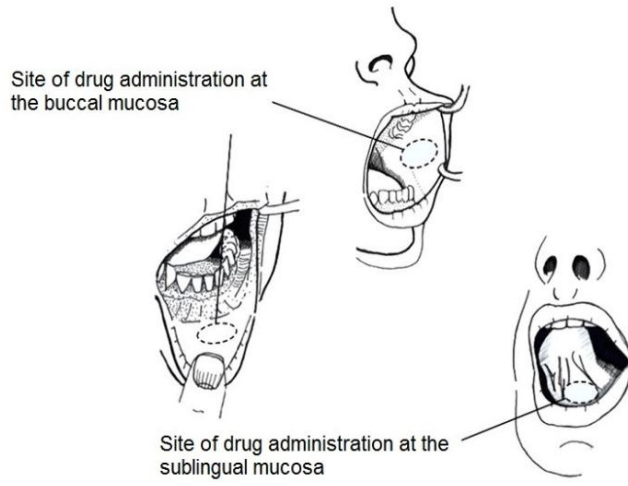
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1135 **Fig. 9**

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Tables

Table 1. Properties and key findings of representative polymers used for preparation of thin film formulations

Polymer	Properties	Key findings	References
Hydroxypropyl methylcellulose (HPMC)	<ul style="list-style-type: none"> White, creamy, odorless, and tasteless powder Mw 10,000–1,500,000 Soluble in cold water, but insoluble in chloroform and ethanol Viscosity (η) 3–100,000 mPa·s Non-ionic polymer with moderate mucoadhesive properties Solutions are stable at pH 3.0 to 11.0 	<ul style="list-style-type: none"> Film forming ability at 2–20% concentrations Generally used for controlled and/or delayed release of the drug substance Initial burst drug release followed by slow or sustained drug release diffusion observed in buccal bioadhesive system of nicotine hydrogen tartrate 	[3, 11, 17, 36]
Carboxymethyl cellulose (CMC)	<ul style="list-style-type: none"> White, odorless powder Mw 90,000–700,000 Easily dispersed in water to form a clear or colloidal solution. η 5–13,000 mPa·s (1% aqueous solution) High swelling properties Good bioadhesive strength 	<ul style="list-style-type: none"> Improved the residence time of HPC and sodium alginate films Good compatibility with starch forming single-phase polymeric matrix films with improved mechanical and barrier properties The enzymatically modified CMC has good film forming property 	[3, 11, 17, 36]
Hydroxypropyl cellulose (HPC)	<ul style="list-style-type: none"> White to slightly yellow colored, odorless, inert and tasteless powder Mw 50,000–1,250,000 Soluble in cold and hot polar organic solvents such as absolute ethanol, methanol, isopropyl alcohol and propylene glycol η 75–6500 mPa·s depending upon the polymer grade Moderate mucoadhesive properties 	<ul style="list-style-type: none"> Used to replace synthetic polymers or HPMC in a polymer matrix with modified starch to improve solubility It has a good film forming property and 5% (w/w) solution is generally used for film coating Zero-order release kinetics of lidocaine and clotrimazole associated with erosion square-root of time release kinetics of lidocaine 	[3, 11, 17, 36]
Poly (vinyl pyrrolidone) (PVP)	<ul style="list-style-type: none"> Wide range of solubility Non-ionic High swelling properties Used as co-adjuvant to increase mucoadhesion 	<ul style="list-style-type: none"> Blending of PVP with PVA and HPMC improve film forming ability Blended with ethyl cellulose and HPC produce films with increased flexibility, softer and tougher properties Different ratios of PVP-alginate blends can be used to design drug controlled release As film-forming polymer exhibited non-Fickian release of ketorolac and progesterone 	[3, 11]
Poly (vinyl alcohol) (PVA)	<ul style="list-style-type: none"> White to cream-colored granular powder Mw 20,000-200,000 	<ul style="list-style-type: none"> Very flexible films Mainly used in ophthalmic polymeric preparations at 	[3]

	<ul style="list-style-type: none"> • Water soluble synthetic polymer • Non-ionic polymer • Moderate mucoadhesive properties 	<ul style="list-style-type: none"> • concentration 3-5% • Higher elongation at break values 	
Poly (ethylene oxide) (PEO)	<ul style="list-style-type: none"> • Non-ionic polymer • High mucoadhesion with high molecular weight 	<ul style="list-style-type: none"> • Optimization of tear resistance, dissolution rate, and adhesion tendencies of film by combining low Mw PEO, with a higher Mw PEO and/or with cellulose • Films with good resistance to tearing, minimal or no curling • Pleasant mouth feeling with no sticky or highly viscous gel formation 	[3, 11]
Pullulan	<ul style="list-style-type: none"> • White, odorless, and tasteless powder • Mw 8000–2,000,000 • Soluble in hot as well as cold water • η 100–180 mm²/s (10% aqueous solution at 30 °C) • Contain > 6% w/w of moisture. 	<ul style="list-style-type: none"> • Blending with sodium alginate and/or CMC, may synergistically enhance the properties of the film. • Pullulan — HPMC films have improved thermal and mechanical properties. • 5–25% (w/w) solution forms flexible films • Stable film with less permeability to oxygen 	[3, 17]
Pectin	<ul style="list-style-type: none"> • A yellowish white, odorless powder with mucilaginous taste • Mw 30,000–100,000 • Soluble in water but insoluble in most of the organic solvents • Strong mucoadhesive properties 	<ul style="list-style-type: none"> • Not very useful for fast dissolving films, but modified pectins yielded films with fast dissolution rates • Good film forming capacity at low temperature • Brittle and do not have a clear plastic deformation. 	[3, 17]
Chitosan	<ul style="list-style-type: none"> • White or creamy powder or flakes, and odorless • Obtained after partial deacetylation of chitin • Biocompatible and biodegradable • Sparingly soluble in water; practically insoluble in ethanol (95%), other organic solvents, and neutral or alkali solutions at pH above approximately 6.5 	<ul style="list-style-type: none"> • Excellent film forming ability • Chitosan enhance the transport of polar drugs across epithelial surfaces • Possesses cell-binding activity due to polymer cationic polyelectrolyte structure that binds to the negative charge of the cell surface 	[11, 36]
Sodium alginate	<ul style="list-style-type: none"> • Occurs as a white or buff powder, which is odorless and tasteless • Purified carbohydrate product extracted from brown seaweed by the use of dilute alkali • Insoluble in other organic solvents and acids where the pH of the resulting solution falls below 3.0 • η 20–400 Cps (1% aqueous solution) • Anionic with high mucoadhesive properties • Safe, biodegradable and non-allergenic • Rapid swelling and dissolution in water 	<ul style="list-style-type: none"> • Used as immobilization matrices for cells and enzymes, controlled release of bioactive substances • Excellent gel and film forming properties • Compatible with most water-soluble thickeners and resins 	[11, 36]

- Carrageenan**
- An anionic polysaccharide, extracted from the red seaweed *Chondrus crispus*
 - Three structural types exist: Iota, Kappa, and Lambda, differing in solubility and rheology
 - The sodium form of all three types is soluble in both cold and hot water
 - The best solution stability occurs in the pH 6 to 10
 - Moderate mucoadhesive properties
- Gelatin**
- A light amber to faintly yellow colored powder
 - Mw 15,000–250,000
 - Soluble in glycerin, acid, alkali and hot water
 - η 4.3–4.7 mPa s (6.67% (w/v) aqueous solution at 60 °C)
 - Moisture content 9–11% (w/w)
- Potential to act as protein/peptide stabilizer by steric stabilization [6, 11, 36]
 - It is compatible with most nonionic and anionic water soluble thickeners
 - Solutions are susceptible to shear and heat degradation
- It has a very good film forming ability [17]
 - Useable for preparation of sterile film, ophthalmic film, and sterile sponge

Table 2. List of commercialized thin films for drug delivery

Company	Brand name	Type of formulation	References
Labtec Pharma	Zolmitriptan Rapidfilm®	Zolmitriptan oral disintegrating films (ODF)	
BioAlliance Pharma	Setofilm®	Ondansetron ODF	[21]
MonoSol Rx and KemPharm	KP106	D-amphetamine ODF	
BioDelivery Sciences International	Onsolis™	Fentanyl buccal soluble films	[11]
Labtec Pharma	RapidFilm®	Ondansetron and donepezil ODF	[2]
Novartis	Triaminic Thin Strips	Phenylephrine and diphenhydramine ODF	
MonoSol Rx	Suboxone®	Buprenorphine and naloxone (sublingual film)	
C.B. Fleet	Pedia-Lax™ Quick Dissolve Strip	Sennosides ODF	[55]
Novartis Consumer Healthcare	Gas-X Thin Strips	Simethicone (sublingual film)	
Pfizer	Sudafed PE quick dissolve strips	Phenylephrine ODF	

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Table 3. List of drugs used in ocular films

Active agent in ocular film	References
Acetazolamide	[102]
Timolol maleate	[103]
Ofloxacin	[104]
Dorzolamide hydrochloride	[105]
Levofloxacin	[78, 106]
Naphazoline HCl	[107]
Natamycin	[108]

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Comment [A1]: Author: There are two table 3 captions were provided in the manuscript and this has been retained. Please check and confirm it is correct.